

Evaluation of reproductive parameters of vas deferens sperms in Caucasian snake (*Gloydius halys caucasicus*)

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Article Info	Abstract
<p>Article history:</p> <p>Received: 27 August 2011 Accepted: 18 January 2012 Available online: 15 June 2012</p> <p>Key words:</p> <p>Snake Sperm Motility Vitality Morphology</p>	<p>Reproductive parameters evaluation is considered as helpful tool for gene bank formation in ecological and economically important animals species. <i>Gloydius halys caucasicus</i> is venomous, viviparous pit viper of northwest of Iran. In this research, the spermatid reproductive parameters of this taxon were studied. Twenty six male snakes were collected from Takht-e-Soleiman region between September and October, 2010. Findings revealed that male snakes with body length of 45.07 ± 2.83 cm and body weight of 51.50 ± 10.42 g, and right and left gonads volume of 0.12 ± 0.03 mL and 0.06 ± 0.01 mL are mature ones and sperms concentration in first, middle and final regions of vas deferens duct were, $22.30 \pm 19.34 \times 10^6$ mL⁻¹, $30.34 \pm 11.55 \times 10^6$ mL⁻¹, and $37.65 \pm 16.46 \times 10^6$ mL⁻¹, respectively. The sperms motility at three regions of duct were 60.53%, 62.07%, and 60.00% and percentage of immotile sperms in these regions were 39.46%, 37.92%, and 39.84%, respectively. Percentage of morphologically normal sperms was $69.23 \pm 10.57\%$ and abnormal sperms was $30.76 \pm 10.57\%$; including $12.69 \pm 5.25\%$ spiral tailed, $7.33 \pm 4.37\%$ coiled tailed and $4.16 \pm 2.51\%$ folded tailed sperms. Percentage of live sperms in the first, middle and final regions of duct were $55.76 \pm 10.77\%$, $58.84 \pm 12.77\%$, and $57.69 \pm 9.91\%$, respectively and percentage of dead sperm in these regions were $44.23 \pm 10.77\%$, $41.15 \pm 12.77\%$, and $42.30 \pm 9.91\%$, respectively. Results suggested, mature sperms with acceptable reproductive quality could be collected from <i>Gloydius halys caucasicus</i> snake of Iran between September and October.</p> <p>© 2012 Urmia University. All rights reserved.</p>

ارزیابی پارامترهای تولید مثلی اسپرم های مجرای دفران در مار قفقازی (*Gloydius halys caucasicus*)

چکیده

ارزیابی پارامترهای تولیدمثلی از ابزارهای کارآمد برای ایجاد بانک ژن در گونه های جانوری که اهمیت زیست محیطی و اقتصادی دارند محسوب می شود. مار قفقازی (*Gloydius halys caucasicus*) که از مارهای سمی شمال غربی ایران است جزو افعی حفره دار و زنده زا می باشد. در این تحقیق پارامترهای باروری اسپرمی در این تاکسون مورد مطالعه قرار گرفت. ۲۶ حلقه مار نر از منطقه تخت سلیمان در شهر یور تا مهر ماه سال ۱۳۸۹ جمع آوری شد. نتایج نشان می دهند وزن متوسط جانور 51.50 ± 10.42 گرم، میانگین طول بدن از پوزه تا منفذ کلوآک 45.07 ± 2.83 سانتی متر، میانگین طول دم 61.07 ± 0.85 سانتی متر بود. میانگین وزن بیضه های راست و چپ به ترتیب 16.01 ± 17.02 و 9.30 ± 17.07 گرم و میانگین حجم بیضه های راست و چپ به ترتیب 0.12 ± 0.11 و 0.06 ± 0.01 میلی لیتر و میانگین طول مجراهای تولید مثلی راست و چپ به ترتیب 1.21 ± 1.30 و 1.00 ± 1.14 سانتی متر بود. میانگین غلظت اسپرم در سه قسمت اولیه، میانی و انتهایی مجرای دفران به ترتیب 22.30 ± 19.34 ، 30.34 ± 11.55 و $37.65 \pm 16.46 \times 10^6$ در هر میلی لیتر بود. میزان تحرک اسپرم در ابتدای مجرا $55.76 \pm 10.77\%$ ، میانه مجرا $58.84 \pm 12.77\%$ و انتهایی مجرای دفران $57.69 \pm 9.91\%$ و درصد اسپرم های غیرمتحرک در ابتدای مجرا 39.46% ، میانه مجرا 37.92% و انتهایی مجرای دفران 39.84% و درصد اسپرم های غیرنرمال شامل دم مارپیچ $12.69 \pm 5.25\%$ و دم حلقه ای $7.33 \pm 4.37\%$ و درصد اسپرم های زنده در ابتدا، میانه مجرا، $55.76 \pm 10.77\%$ ، میانگین مجرای دفران $58.84 \pm 12.77\%$ و $57.69 \pm 9.91\%$ و درصد اسپرم های مرده در ابتدا، میانه و انتهایی مجرای دفران به ترتیب $44.23 \pm 10.77\%$ ، $41.15 \pm 12.77\%$ و $42.30 \pm 9.91\%$ بود. براساس یافته ها پیشنهاد می شود اسپرم های بالغ با کیفیت تولید مثلی قابل قبول را می توان بین ماه های شهریور تا مهر از مارسمی قفقازی ایران جمع آوری نمود.

واژه های کلیدی: مار، اسپرم، تحرک، ماندگاری، مورفولوژی

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Introduction

The Caucasian pit viper, *Gloydius halys caucasicus*, is a venomous and viviparous species belonging to the Crotalidae family. It is widely distributed in different parts of Iran¹⁻³ and its habitat is in provinces of Tehran, Gilan, Mazandaran, Golestan, North Khorasan, Razavi Khorasan and Semnan.^{1,2} Often in temperate zone the mating season of snakes is temporally dissociated from the time of fertilization. Similarly, in males, the mating season is often temporally dissociated from spermatogenesis. In temperate zone pit vipers of North America, estrus, the time when females signal that they are receptive to males, occurs at some time during vitellogenesis. In these pit vipers, vitellogenesis initiates in the late summer or fall. In tropical pit vipers the vitellogenic cycle is continuous (no winter pause) and the mating season occurs at some time during vitellogenesis.⁴ Snakes display considerable variability in reproductive traits among species^{5,6} exhibiting a wide range of mating systems and male and female reproductive behaviors.^{7,8} Saint-Girons has described four patterns of male reproduction.⁹ First, the postnuptial (dissociated) or estival type, which occurs in many temperate and subtropical snake species, is a pattern in which male snake undergo spermatogenesis during the summer, sperms are stored over the winter in the male vasa deferentia and sometimes also in female oviducts (if fall mating occurs), and the principal mating season occurs in spring. This postnuptial pattern of spermatogenesis is predominant among temperate colubrids and crotalines. Second is the prenuptial (associated) or vernal type, which typically occurs in warm climates. In this type, spermatogenesis begins in fall and is completed by the following spring or early summer, when mating occurs. Third is the mixed type characterized by spermatogenesis beginning in spring and being completed one year later, can be one (spring) or two (spring and fall) mating seasons. As with the postnuptial pattern, the mixed type typically occurs in temperate and subtropical species. Finally, the continuous type describes species in environments where there is little seasonal variation in temperature, (e.g., most tropical areas). As the name implies, species that express continuous male reproduction exhibit spermatogenesis and mating behavior throughout the year. Although distinct in definition, these four types of male reproduction in snakes should be viewed as noteworthy points along a cline from distinctly seasonal (estival spermatogenesis) to aseasonal (continuous spermatogenesis), exceptions certainly exist.⁹ Few studies have focused on snake sperm motility^{10, 11} and all of them have estimated this parameter in a subjective manner. Only one study has thoroughly described the basic sperm dynamic parameters, velocity and motility of a snake species.¹² It is widely accepted that the regulation of sperm motility and fertilizing ability depends on the inter-

action of several factors, among which temperature is of vital importance.¹³ Still, the effect of temperature on sperm dynamic parameters in ectotherms in genera^{14,15} and reptiles in particular^{10, 16} remains poorly understood due to the lack of consistent evidence. In a research done in Brazil it was shown that the female *Gloydius halys caucasicus* ovulated in the beginning of spring and mated with male snakes. At the time of ovulation and pregnancy, level of progesterone increased and serum level remained high. Pregnancy lasted for four months, until the end of summer. The average number of neonates was 3 to 9.^{17, 18} Thus spermatogenesis began from the end of spring in Caucasian snake, and its climax was in the summer, and continued until autumn.¹⁹ Summer spermatogenesis takes place more often in snake living in temperate and cold regions.²⁰ In this research Caucasian viper, (*Gloydius halys caucasicus*), of the Iran was studied. It is among dangerous and poisonous snakes and used for production of polyvalence antitoxin for snake bites. The enzymes existing in its poison are also of the pharmaceutical importance.^{21, 22}

This research contributes to conservation of special and rare species in the country and helps to identify and make optimal use of this reptile in the laboratory research.²¹ More comprehensive scientific information about snakes of Iran could be found in the books: "Snakes of Iran" by Mahmud Latifi,^{1,2} "Mar Shenakht" by Farzanpey²³ and "Encyclopedia of Hayat- e -Vahsh- e -Iran" by Ibrahimi.²⁴ There is no information about spermatogenic parameters of snakes in Iran as yet. In our research the reproductive parameters of sperm in Iran's Caucasian snake have been studied for the first time.

Materials and Methods

Twenty six Caucasian male snakes were collected from Takht-e-Soleiman (average annual temperature of 9.5°C, maximum of 27.6 °C, and minimum of -11.3 °C). All procedures were carried out in accordance with ISIRI 7216-2 animal ethical guidelines.²⁵ The sperm samples were collected from September to October (in viperidae, vasa deferentia contain sperm from February to October).²⁶ The snakes were kept in cages (length 150 cm, width 40-60 cm, height 70 cm) with a temperature of 20-30 °C, 10/14 hr light/dark and fed every second week. Snake was anesthetized, using subcutaneous injection of 1% lidocaine (15 mg kg⁻¹, DarouPakhsh, Iran) around the cloacae. The anesthetic was diluted to a total volume of 1.0 mL in normal saline. The total volume of anesthetic was divided between four injection sites (0.25 mL per site) anterior to the cloacae.¹¹ Snakes were sacrificed by injection of 1.0 mL ethanol (96%, Bidestan Co., Iran) to each optical cavity. The body length of snakes was measured from the tip of their snout to the vent by ordinary measuring tape.²⁷ The testes and vas deferens

were removed, then the dimensions of their right and left gonads including lengths, widths and heights were measured by caliper and volumes were calculated by Cha formula.²⁸ The vas deferens was divided into three parts of first, middle, and final. Each part was separately placed in Petri dishes, and cut into pieces by sterilized blade and scissors, and immediately kept in vials in 1mL PBS for 45 minutes. During this period the gonads were weighted (Scale No. 11327, Japan). Sperm motility, morphology, vitality, and concentration were studied. The sperm cells were counted using a hemocytometer under 400× and concentration of sperm was calculated according to Rashidi *et al.*²⁹ In order to study the sperm motility, four grades were considered: A (quick progressive in straight paths), B (slow progressive in straight or not straight paths), C (motile in place), D (immotile).³⁰⁻³² Morphology was evaluated by observing 100 sperm cells under microscope at 1000×, and finally the survival period of sperm cells was measured in laboratory temperature of 23 ± 2 °C.

Statistical analyses were performed by Student *t*-test for comparison of two values at $P < 0.0001$ and one way ANOVA where three values were compared at $P < 0.05$.

Results

Average weight of the animals, body length, length of tail, details of testes (weight, volume and length) the length of right and left reproductive ducts were measured and have been shown in Table 1. Our results indicated that the right testis was bigger than left one and their average volume, weight and length (Table 1) were significantly different at $P < 0.0001$ and $P = 0.001$, respectively.

Table 1. Some morphometric specifications of *Gloydius halys caucasicus* snake (Mean ± SD).

Variable	Value
Body length (from snout to vent) (cm)	45.07 ± 2.83
Length of tail (cm)	6.07 ± 0.85
Length of left vas deferens from testis to vent (cm)	15.30 ± 1.21
Length of right vas deferens from testis to vent (cm)	11.74 ± 1.003
Weight of right testis (mg)	123.07 ± 16.01
Weight of left testis (mg)	93.07 ± 17.02
Volume of right testis (mL)	0.12 ± 0.03
Volume of left testis (mL)	0.06 ± 0.01
Length of right testis (mm)	21.77 ± 3.70
Length of left testis (mm)	16.69 ± 3.95

Length of left vas deferens duct was longer than right one at $P < 0.0001$. Sperms concentration and vitality in first, middle and final parts of vas deferens, motility, morphology of sperms have been presented in Table 2. Sperm concentrations were not significantly different in three parts of duct ($P > 0.05$), percentages of live and dead sperms were not statistically different in three parts of duct at $P > 0.05$, percentages of motile & immotile sperms (Table 3) differences in three parts of duct were not considered significant ($P > 0.05$).

Table 2. Concentration, vitality, survival and morphology of sperms in *Gloydius halys caucasicus* snake (Mean ± SD).

Variable	Value
Sperm concentration in the first region ($\times 10^6$ mL ⁻¹)	22.30 ± 19.34
Sperm concentration in the middle region ($\times 10^6$ mL ⁻¹)	30.34 ± 11.55
Sperm concentration in the final region ($\times 10^6$ mL ⁻¹)	37.65 ± 16.46
Live sperms (first region of duct) (%)	55.76 ± 10.77
Live sperms (middle region of duct) (%)	58.84 ± 12.77
Live sperms (final region of duct) (%)	57.69 ± 9.91
Dead sperms (first region of duct) (%)	44.23 ± 10.77
Dead sperms (middle region of duct) (%)	41.15 ± 12.77
Dead sperms (final region of duct) (%)	42.30 ± 9.91
Sperms survival period (hours)	6.00 ± 2.00
Normal sperms (%)	69.23 ± 10.57
Abnormal sperms (%)	30.76 ± 10.57
Abnormal spiral sperms (%)	12.69 ± 5.25
Abnormal coiled sperms (%)	7.33 ± 4.37
Abnormal short-tailed sperms (%)	8.00 ± 6.99
Abnormal folded tailed sperms (%)	4.16 ± 2.51

Table 3. Sperm motility grades in *Gloydius halys caucasicus* snake (ranges of values).

Motile sperm in first region of duct (%)	60.53 (45-84)
Grade A	4.07 (0-13)
Grade B	6.30 (1-13)
Grade C	50.15 (39-67)
Grade D (immotile sperms) (%)	39.46 (16.55)
Motile sperm in middle region of duct (%)	62.07 (43-86)
Grade A	7.46 (0-33)
Grade B	5.53 (3-14)
Grade C	49.07 (38-68)
Grade D (immotile sperms) (%)	37.92 (14-57)
Motile sperm in final region of duct (%)	60.00 (43-75)
Grade A	8.00 (0-23)
Grade B	4.53 (0-10)
Grade C	47.46 (36-66)
Grade D (immotile sperms) (%)	39.84 (25-57)

Discussion

This study represents basic values of the reproductive potential of Iranian *Gloydius halys caucasicus* which is an important step in the development of assisted reproduction in snakes. The concentration of sperm obtained in our experiment were within ranges of results obtained for checkered garter and *Crotalus durissus terrificus* snakes using similar methods.³³⁻³⁵ Morphologically, 75.70% normal sperm cells reported in corn snakes' ejaculates,¹⁰ was different from our results (69.23%). The percentage of normal morphology in corn snakes is comparable to that of the ram, bull, boar, stallion, and buck.³⁶ In mammals, a small range of morphologic abnormalities are considered normal in a healthy animal. Among the commonly measured semen parameters, morphologic abnormalities have the greatest negative correlation to fertility of farm animals, and heat stress is one of the main causes of sperm abnormalities in these animals. The effect of heat stress on reptile spermatozoa has not yet been assessed. Reptiles are ectotherms and depend on their environmental temperature to regulate

their core temperature. It would not be unexpected for a *Gloydius halys caucasicus* snake to experience a 2.7-5.5 °C difference in body temperature in a given day. Although it has not been evaluated, one might expect that semen production would diminish and sperm abnormalities would increase in snakes that experience a large drop in body temperature. Genetics and exogenous drugs have also been associated with sperm cell abnormalities in higher vertebrates.

In normal fertile individuals up to 50% of sperm cells can have morphological defects (up to 70% according to WHO³¹ criteria and up to 86% according to strict criteria).³⁵ Sperm morphology gives information for the function of the reproductive tract and is a predictor of animal's fertility potential. Generally it seems that Caucasian snakes in Iran with SVL 45.07 ± 2.83 cm and testis volume of right 0.12 ± 0.03 mL and left, 0.06 ± 0.01 mL, is mature,³⁶ like other snakes in cold and mild regions, have got summer spermatogenesis.¹⁹

The findings represent the qualitative and quantitative values of sperms of Iran's Caucasian snake from late September to early October, however in order to obtain higher quality sperms, more work is needed to be carried out.

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References

1. Latifi M. Snakes of Iran. 3rd ed. Tehran: Department of the Environment 2000; 36.
2. Latifi M. Snakes of Iran. 1st ed. Tehran: Department of the Environment 1991; 478.
3. Rastegar-pouyani N, Kami HG. Annotated checklist of amphibians and reptiles of Iran. *Iranian Journal Animal Biosystematics* 2008; 4(1): 43-66.
4. Aldridge RD, Duvall D. Evolution of the mating season in the pitvipers of North America. *Herpetol Monogr* 2002; 16(1):1-25.
5. Hartmann MT, Marques OAV, Almeida-Santos SM. Reproductive biology of the southern Brazilian pitviper *Bothrops neuwiedi pubescens* (Serpents, Viperidae). *Amphibian Reptile* 2004; 25(1): 77- 85.
6. Zug GR, Vitt LJ, Caldwell JP. *Herpetology: An Introductory Biology of Amphibians and Reptiles*. San Diego: Academic Press, 2001; 1213-1223.
7. Duvall D, Arnold SJ, Schuett GW. Pitviper mating systems: Ecological potential, sexual selection and micro-evolution. In: Campbell JA, Jr. Brodie ED. Eds. *The Biology of the Pitvipers*. Tyler: Selva Press 1992; 321-336.
8. Tourmente M, Gomendio M, Roldan ERS, et al. Sperm competition and reproductive mode influence sperm dimensions and structure among snakes. *Evolution* 2009; 63(10): 2513-2524.
9. Saint-Girons H. Reproductive cycles of male snakes and their relationships with climate and female reproductive cycles. *Herpetologica* 1982; 38: 5-16.
10. Fahrig MB, Mitchell MA, Eilts BE, et al. Characterization and cooled storage of semen from corn snakes (*Elapha Guttata*) *J Zoo Wildl Med* 2007; 38(1): 7-12.
11. Zacariotti RL, Zuge RM, Grego KF, et al. Semen collection and evaluation in the Brazilian rattlesnake (*Crotalus durissus terrificus*): preliminary results. In *Proceedings: The 2nd International Symposium on Assisted Reproductive Technology for the Conservation and Genetic Management of Wildlife*. Omaha, USA, 2002; 26: 155-160.
12. Tourmente M, Cardozo GA, Guidobaldi HA, et al. Sperm motility parameters to evaluate the seminal quality of *Boa constrictor occidentalis*, a threatened snake species. *Res Vet Sci* 2007; 82(1): 93-98.
13. Ashizawa K, Wishart G. Sperm motility in the female reproductive tract. In *Proceedings: The 9th European Poultry Congress*, Glasgow, UK1994; 173-176.
14. Costanzo J P, Mugnano JA, Wehrheim HM, et al. Osmotic and freezing tolerance in spermatozoa of freeze-tolerant and -intolerant frogs. *Am J Physiol Regul Integr Comp Physiol* 1998; 275:713-719.
15. Johnson SL, Yund PO. Remarkable longevity of dilute sperm in a free-spawning colonial ascidian. *Biol Bull* 2004; 206:144-151.
16. Gist D, Turner T, Congdon J. Chemical and thermal effects on the viability and motility of spermatozoa from the turtle epididymus. *J Reprod Fertil* 2000; 119: 271-277.
17. Almeida-Santos SM, Laporta-Ferreira I L, Antoniazzi MM, et al. Sperm storage in males of the snake *Crotalus durissus terrificus* (Crotalinae: Viperidae) in South-eastern Brazil. *Comp Biochem Physiol A Mol Integr Physiol* 2004; 139(2): 169-179.
18. Siegel DS, Sever DM. Seasonal variation in the oviduct of female *Agkistrodon piscivorus* (Reptilia: Squamata): An ultrastructural investigation. *J Morphol* 2008; 269: 980-997.
19. Almeida- Santos SM, Abdalla FMF, Silveira PF, et al. Reproductive cycle of the Neotropical *Crotalus durissus terrificus*: I. Seasonal levels and interplay between steroid hormones and vasotocinas. *Gen Comp Endocr* 2004; 139(2): 143-150.
20. Feriche M, Pleguezuelos JM, Santos X. Reproductive ecology of Montpellier Snake, *Malpolon monspessulanus* (Colubridae), and comparison with other sympatric Colubrids in Iberian peninsula. *Copeia* 2008; 2: 279-285.

21. Ghorbanpur M, Zare Mirakabadi A, Zokaee F, et al. Purification and partial characterization of a coagulant serine protease from the venom of the Iranian snake *Agkistradon halys*. *J Venom Anim Toxins incl Trop Dis* 2009; 15(3): 411- 423.
22. Zare Z, Eimani H, Mohammadi M, et al. The effect of orally administered L-carnitine on testis tissue, sperm parameters and daily sperm production in adult mice. *Yachted* 2010; 11(4): 382-389.
23. Farzanpey, R. *Maar shenakht*. 1st ed. Tehran: Academic publication Center 1990; 272.
24. Ibrahim M, Hosseinizavarie F, Rajabizadeh M. et al. *Encyclopedia of Hayate-Vahshe-Iran*, 1st ed. Tehran: Talaei Publishers 2011; 355.
25. Institute of standards and industrial research of Iran 2008. Biological evaluation of medical devices- Part2: Animal Welfare Requirements. ISIRI 7216-2.
26. Goldberg SR. Testicular Cycle of the Western Diamondback Rattlesnake, *Crotalus atrox* (Serpentes: Viperidae), from Arizona. *Bull Md Herpetol Soc* 2007; 43(3): 103-107.
27. Blouin-Demer G. Precision and accuracy of body-size measurements in a constricting, large-bodied snake (*Elaphe obsoleta*). *Herpetol. Rev*, 2003, 34(4), 320-323.
28. Cha MH, Ahn BC, Kim YS. Inaccuracy in ultrasonographic measurement of the T children. *Korean J Urol* 2006; 47(8): 866-869.
29. Rashidi I, Movahedin M, Tiraihi T. The effects of pentoxifylline on mouse epididymal sperm parameters, fertilization and cleavage rates after short time preservation. *IJRM* 2004; 2(2): 51-57.
30. Dada R, Gupta NP, Kucheria K. Deterioration of sperm morphology in men exposed to high temperature. *J Anat Soc India* 2001; 5(2):107-111.
31. World Health Organization. WHO Laboratory Manual for the Examination of Human Semen and Semen-Cervical Mucus Interaction. 3rd ed. Cambridge, United Kingdom: Cambridge University Press 1992:1-128.
21. Samour JH. Recent advances in artificial breeding techniques in birds and reptiles. *Int Zoo Yearb* 1986; 24/25:143-148.
22. Mengden AG, Platz CG, Hubbard R, et al. Semen collection, freezing and artificial insemination in snakes. In: Murphy JB, Collins JT. Eds. Contributions to herpetology reproductive biology and diseases of captive reptiles. St. Louis: St. Louis University press 1980; 71-78.
23. Quinn H, Blasedel T, Platz CC. Successful artificial insemination in the checkered garter snake. *Int Zoo Yearb* 1989; 28: 177-183.
24. Ax RL, Dally M, Didion BA, et al. Semen evaluation. In: Hafez B, Hafez ESE. Eds. *Reproduction in Farm Animals*. 7th ed. Philidelphia: Lippincott, Williams & Wilkins, 2000; 365-375.
25. Wangkulangkul S. Sexual size dimorphism and reproductive cycle of the little file snake *acrochordus granulatus* in phangngabay, Thailand *Science* 2005; 31: 257-263.