

The comparison of digestibility of treated sugarcane tops silage by bacteria or whole microorganisms of Holstein cow and buffalo rumen

Afroz Sharifi¹, Morteza Chaji^{2*}, Tahereh Mohammadabadi²

¹ Graduate Student of Animal Nutrition, Ramin Agriculture and Natural Resources University of Khuzestan, Ahvaz, Iran; ² Department of Animal Science, Ramin Agriculture and Natural Resources University of Khuzestan, Ahvaz, Iran.

Article Info	Abstract
<p>Article history:</p> <p>Received: 25 November 2013 Accepted: 20 May 2014 Available online: 15 September 2016</p> <p>Key words:</p> <p>Bacteria Buffalo Molasses Sulfuric acid Urea</p>	<p>The aim of this study was to evaluate the effects of adding sulfuric acid to sugarcane tops silage on rumen bacteria and whole rumen microorganisms (WRM) and compare the digestibility of sugarcane tops treated with different amount of urea, molasses and sulfuric acid between Holstein cow and Khuzestan buffalo. Regardless of the type of the treatment, potential of gas production (B) by cow WRM (130.670 mL) was more than buffalo (104.060 mL) ($p < 0.05$), but the rate of gas production (C) by buffalo WRM was greater than cow (0.021 and 0.014 mL per hr, respectively) ($p < 0.05$). The C in treatment containing only 2.40% sulfuric acid (0.033 mL per hr) was significantly highest ($p < 0.05$). Regardless of the type of the treatment, the B by cow rumen bacteria (75.040 mL) was more than buffalo (67.150 mL), ($p < 0.05$), while the C by rumen bacteria of buffalo (0.030 mL per hr) was more than cow (0.017 mL per hr), ($p < 0.05$). Regardless of the type of the animal, the B coefficient of rumen bacteria in treatment only containing 2.40% sulfuric acid was higher than control ($p < 0.05$). Therefore, the addition of sulfuric acid not only had no negative effect on microorganisms particularly bacteria, but also probably due to present of sulfur in acid, had positive effect on nutrients digestibility, and growth of microorganisms. The digestibility of sugarcane tops silage treated by cow rumen bacteria and whole microorganisms was higher than buffalo.</p> <p>© 2016 Urmia University. All rights reserved.</p>

مقایسه قابلیت هضم سیلاژ سرشاخه های نیشکر عمل آوری شده توسط باکتری ها یا کل میکروارگانسیم های شکمبه گاو هلشتاین و گاومیش

چکیده

هدف این مطالعه بررسی اثرات افزودن اسید سولفوریک به سیلاژ سر شاخه های نیشکر روی باکتری ها و کل میکروارگانسیم های شکمبه و مقایسه قابلیت هضم سرشاخه های نیشکر تیمار شده با مقادیر مختلف اوره، ملاس و اسید سولفوریک بین گاو هلشتاین و گاومیش خوزستان بود. صرف نظر از نوع تیمار، پتانسیل تولید گاز توسط کل میکروارگانسیم های شکمبه گاو (۱۳۰/۶۷۰ میلی لیتر) بیشتر از گاومیش (۱۰۴/۰۶۰ میلی لیتر) بود ($p < 0.05$) اما میزان تولید گاز توسط کل میکروارگانسیم های شکمبه گاومیش بیشتر از گاو بود (به ترتیب، ۰/۰۲۱ و ۰/۰۱۴ میلی لیتر بر ساعت)، ($p < 0.05$). میزان تولید گاز تیمار حاوی تنها ۲/۴۰ درصد اسید سولفوریک (۰/۰۳۳ میلی لیتر بر ساعت) به شکل معنی داری بالاترین مقدار را داشت ($p < 0.05$). صرف نظر از نوع تیمار، پتانسیل تولید گاز در حضور باکتری های شکمبه گاو (۷۵/۰۴۰ میلی لیتر) بیشتر از گاومیش (۶۷/۱۵۰ میلی لیتر) بود ($p < 0.05$), در حالی که میزان تولید گاز توسط باکتری های شکمبه گاومیش (۰/۰۳۰ میلی لیتر بر ساعت) بیشتر از گاو (۰/۰۱۷ میلی لیتر بر ساعت) بود ($p < 0.05$). صرف نظر از نوع حیوان، پتانسیل تولید گاز توسط باکتری های شکمبه در تیمار حاوی تنها ۲/۴۰ درصد اسید سولفوریک بالاتر از شاهد بود ($p < 0.05$). بنابراین، افزودن اسید سولفوریک نه تنها اثر منفی بر میکروارگانسیم ها به ویژه باکتری ها نداشت، بلکه احتمالاً به واسطه وجود گوگرد در اسید، دارای اثر مثبت بر قابلیت هضم مواد مغذی و رشد میکروارگانسیم ها نیز بود. در مقایسه با گاومیش، قابلیت هضم سیلاژ سر شاخه های نیشکر تیمار شده توسط باکتری ها و کل میکروارگانسیم های شکمبه گاو بیشتر بود.

واژه های کلیدی: اسید سولفوریک، اوره، باکتری ها، گاومیش، ملاس

*Correspondence:

Morteza Chaji. BSc, MSc, PhD
Department of Animal Science, Ramin Agriculture and Natural Resources University of Khuzestan, Ahvaz, Iran.
E-mail: chaji@ramin.ac.ir

Introduction

Cultivation history of sugarcane in southern Iran is very long, especially in Khouzestan province. In Iran, quantity of sugarcane production is over 6,000,000 tons.¹ Therefore, extraction of sugar from sugarcane produces large quantities of by-products which could be valuable feedstuff for ruminants feeding in dry seasons. Sugarcane tops (SCT) are major by-products of the sugarcane industry which are often left in the field after cane harvest.² Approximately 1.40 million tons of SCT are produced annually in the Khouzestan province. However, the low digestibility and high lignin are considered as main reasons for unsatisfactory performance of animals fed with this roughage.³ Considering limited harvest season, high levels of SCT production and low nutritional value of SCT, using silo to preserve (and use in other seasons) and enrich SCT with some additives could be useful.

Generally, silage additives are edible materials including urea, molasses, bacteria inoculants and acids. Molasses as a cheap carbohydrate source for lactic acid bacteria provides necessary sugar and carbohydrate for fermentation process. Many experiments have proved that molasses increases lactic acid fermentation and reduces silage pH.⁴ Digestibility of SCT crude protein is low,⁵ therefore, using a suitable source of nitrogen, which improves sugarcane nutritional value, is useful. These additives (e.g. urea) are useful when they can be easily fermented by microorganisms.⁶ On the other hand, due to slow reduction in pH, it was reported that using urea lonely, is unable to stop proteolysis, entirely.⁷ Although urea + molasses,^{4,5} or molasses alone,⁴ can improve digestion and storage duration of SCT,⁶ but these additives are not suitable for long term storage of silage.^{4,6} Previously, formic acid and sulfuric acid have been used to decrease the pH of silage.⁸ It was found that rapid acidification of silage is important, and application of acids leads to rapid decrease of pH to below 4 and prevents proteolysis activity.⁸ However, there is conflicting information about capacity of sulfuric acid to lower the pH and probable negative effect of sulfur on activity of rumen bacteria.^{9,10} It has been reported that 0.32% sulfur increases the population of the rumen microorganisms.⁹ But in another study, when sulfur (as sodium sulfate) content of diet was 0.30%, the growth of rumen microbes restrained, and synthesis of microbial protein decreased.¹⁰

The reported populations of microorganism in the rumen of cattle and buffalos have been different.^{11,12} Many factors such as physiological situation, animal age, feeding behavior, level of production, animal health, the nature and relationships between different microbial populations and also external factors such as diet composition, nature of feed, feed frequency, dietary changes, change of seasons and geographical factors can affect the ratio and density of different groups of rumen microorganisms.¹³

Therefore, the aim of the present study was to assess the digestibility and feeding value of acid sulfuric and molasses + urea treated SCT silage by gas test running with different inoculums from Holstein cow and buffalo.

Materials and Methods

This experiment was carried out at Ramin Agriculture and Natural Resources University of Khouzestan, Ahvaz, Iran. Sugarcane tops for ensiling were prepared from Amir Kabir Agro-Industry (Ahvaz, Iran). Sugarcane tops were chopped (3 to 5 cm) and ensiled into plastic bags in triplicate (4 kg). Before ensiling, SCT were mixed with respective treatments well and transferred to bags. Then, they packed massively until no air remains inside the bags. After 120 days, the silos were opened. Experimental treatments were: 1) SCT ensiled without additive (control), 2) SCT ensiled with urea (1.00%) + molasses (3.00%), 3) SCT ensiled with sulfuric acid 2.40% and 4) SCT ensiled with urea (1.00%) + molasses (3.00%) and sulfuric acid (2.40%).

Gas production (GP) experiments were run three times, each run as a repeat. Rumen fluid was collected from two fistulated cattle (weighted 430 ± 12 kg) and buffalo steer (weighted 420 ± 14 kg) before morning feeding. They were fed twice per day with maintenance diet (including alfalfa hay, wheat straw, sugarcane pith, soybean meal, barley, corn, urea, minerals and vitamin materials). Collected rumen liquid was strained through four layers of cheesecloth and mixed with an appropriate volume of artificial saliva.

Gas production of experimental treatments by whole rumen microorganisms (WRM) was measured according Menk and Stingenss method in 100 mL glass syringes containing 300 mg of ground sample, 20 mL of artificial saliva and 10 mL of rumen liquid.¹⁴ The same method was used to determine GP of rumen bacteria, but for isolation of rumen bacteria, after collection and straining of liquor, rumen fluid was centrifuged (1000 rpm for 10 min) for removing protozoa. Then, bacteria were isolated from non-protozoa strained rumen fluid using antifungal agents benomyle (500 mg L^{-1}) and metalaxyle (10 mg L^{-1}).¹⁵ Gas production was measured after 2, 4, 6, 8, 12, 16, 24, 36, 48, 72 and 96 hr. For measurement of cell wall degradability, the syringes contents were carefully filtered (No. 42; Whatman, Pittsburgh, USA) and then, the residues were washed with distilled water into a tube, separately. Then, the residues were dried at 105°C for 12 hr and used to calculate the degradation of samples. Cumulative GP data were fitted to the exponential equation:

$$Y=b(1-e^{-ct})$$

where, b is GP from fermentable fraction (mL), c is GP rate constant (mL per hr), t is the incubation time (hr) and Y is the gas produced at time t .¹⁶

To estimate partitioning factor (PF), which expresses as ratio of truly degraded organic matter to gas produced in incubation periods,¹⁷ at the end of incubation period, the content of each syringe was transferred into Erlenmeyer flask (Schott, Mainz, Germany), mixed with 20 mL neutral detergent fiber solution, boiled for 1 hr and then, filtered, oven-dried at 60 °C for 48 hr (Memmert, Schwabach, Germany) and ashed in furnace at 550 °C for 3 hr (Exciton, Tehran, Iran). Partitioning factor, microbial biomass and actually degradable organic matter were calculated by Makkar method.¹⁸

Data were analyzed by split plot design, main plot was animal species (cattle or buffalo) and subplot included microorganism type (WRM or bacteria). Using general linear model procedures of SAS (Version 9.1; SAS Institute, Cary, USA), Duncan's multiple range tests were used to compare the means.

Results

Gas production by WRM. In buffalo, the highest potential of GP was for SCT ensiled with 2.40 % sulfuric acid (134.83 mL) and in cattle was for control treatment (146.56 mL), ($p < 0.05$). The highest rate of GP in buffalo was for SCT ensiled with sulfuric acid (0.045 mL per hr) and for cattle was in urea + molasses + sulfuric acid (0.021 mL per hr) treatment ($p < 0.05$), (Table 1).

Regardless of the type of the treatment, potential of GP by WRM of cattle was significantly ($p < 0.05$) higher than buffalo (130.670 and 104.060 mL, respectively), but GP rate was significantly ($p < 0.05$) higher in buffalo (0.021 and 0.014 mL per hr, respectively), (Table 1).

Regardless of the type of the animal, the effect of different processing methods on potential of GP of SCT silage was not significant. But, numerically the highest potential and rate of GP (123.940 mL, 0.033 mL per hr, respectively) were in the treatment containing 2.40% sulfuric acid (Table 1).

Gas production by rumen bacteria. The effect of different treatments on GP of SCT silage by rumen bacteria was significant (Table 2). In buffalo, the highest potential of GP was in the treatment containing 2.40% sulfuric acid ($p < 0.05$). In buffalo, GP rate of SCT ensiled with 2.40% sulfuric acid (0.068 mL per hr), and in cattle, GP rate of SCT ensiled with urea + molasses + sulfuric acid (0.025 mL per hr) was the highest amount ($p < 0.05$).

Regardless of the type of the treatment, potential of GP by rumen bacteria of cattle was significantly higher than buffalo (75.040 and 167.150 mL, respectively). Reversely, GP rate in buffalo was higher (0.030 and 0.017 mL per hr, respectively; $p < 0.05$), (Table 2).

Regardless of the type of the animal, the potential of GP by the rumen bacteria for all treatments was higher than control ($p < 0.05$). The rate of GP in treatments containing 2.40% sulfuric acid (0.046 mL per hr) and urea + molasses + sulfuric acid (0.035 mL per hr) was significantly more than control (0.006 mL per hr; $p < 0.05$), (Table 2).

Gas production parameters by WRM. The results showed that the highest microbial biomass was in diet containing urea + molasses + sulfuric acid ($p < 0.05$) for both buffalo (50.30 mg) and cattle (62.75 mg). In buffalo, the highest PF and microbial biomass efficiency were in control (4.30 mg mL⁻¹, 49.00%, respectively). In cattle, the highest PF and microbial biomass efficiency were in

Table 1. Gas production parameters of treated sugarcane tops silage by whole rumen microorganisms of cattle and buffalo.

Treatment	Animal	Urea + Molasses (%)	Sulfuric acid (%)	Potential of gas production (mL)	Gas production rate (mL hr ⁻¹)
1	Buffalo	0.00	0.00	81.000 ^b	0.004 ^d
2		1.00 + 3.00	0.00	89.600 ^b	0.004 ^d
3		0.00	2.40	134.830 ^a	0.045 ^a
4		1.00 + 3.00	2.40	110.130 ^{ab}	0.031 ^b
1	Cattle	0.00	0.00	146.560 ^a	0.006 ^d
2		1.00 + 3.00	0.00	145.180 ^a	0.009 ^d
3		0.00	2.40	113.050 ^{ab}	0.020 ^c
4		1.00 + 3.00	2.40	115.870 ^{ab}	0.021 ^c
SEM				10.980	0.002
<i>p</i> -value				0.0045	0.0001
Regardless of the type of the treatment					
Buffalo				104.060 ^b	0.021 ^a
Cattle				130.170 ^a	0.014 ^b
SEM				5.490	0.0010
<i>p</i> -value				0.0040	0.0001
Regardless of the type of the animal					
1		0.00	0.00	114.120	0.005 ^c
2		1.00 + 3.00	0.00	117.390	0.007 ^c
3		0.00	2.40	123.940	0.033 ^a
4		1.00 + 3.00	2.40	113.000	0.026 ^b
SEM				7.760	0.0014
<i>p</i> -value				0.7540	0.0001

SEM: Standard error of the means; Different superscripts within each column indicate significant differences ($p < 0.05$).

Table 2. Gas production parameters of treated sugarcane tops silage by rumen bacteria of cattle and buffalo.

Treatment	Animal	Urea + Molasses (%)	Sulfuric acid (%)	Potential of gas production (mL)	Gas production rate (mL hr ⁻¹)
1		0.00	0.00	47.740 ^b	0.005 ^{ef}
2	Buffalo	1.00 + 3.00	0.00	71.200 ^a	0.002 ^f
3		0.00	2.40	78.010 ^a	0.068 ^a
4		1.00 + 3.00	2.40	71.640 ^a	0.045 ^b
1		0.00	0.00	72.940 ^a	0.008 ^{de}
2	Cattle	1.00 + 3.00	0.00	73.290 ^a	0.012 ^d
3		0.00	2.40	77.710 ^a	0.023 ^c
4		1.00 + 3.00	2.40	76.230 ^a	0.025 ^c
SEM				4.600	0.001
<i>p</i> -value			0.0070	0.0001	
<i>Regardless of the type of the treatment</i>					
Buffalo				67.150 ^b	0.030 ^a
Cattle				75.040 ^a	0.017 ^b
SEM				2.340	0.000
<i>p</i> -value				0.030	0.000
<i>Regardless of the type of the animal</i>					
1		0.00	0.00	60.340 ^b	0.006 ^c
2		1.00 + 3.00	0.00	72.240 ^a	0.007 ^c
3		0.00	2.40	77.860 ^a	0.046 ^a
4		1.00 + 3.00	2.40	73.940 ^a	0.035 ^b
SEM				3.320	0.0010
<i>p</i> -value				0.0109	0.0001

SEM: Standard error of the means; Different superscripts within each column indicate significant differences ($p < 0.05$).

the treatment containing sulfuric acid (3.48 mg mL⁻¹ and 37.00%, respectively), ($p < 0.05$). The highest true organic matter disappearance (TOMD) was in the treatment containing sulfuric acid (171.50 mg) in buffalo, and in cattle, the highest TOMD was in urea + molasses + sulfuric acid treatment (178.25 mg; $p < 0.05$), (Table 3).

Regardless of the type of the treatment, the PF ($p < 0.05$) in buffalo (3.46 mg mL⁻¹) was higher than cattle (3.21 mg mL⁻¹). However, microbial biomass and TOMD of cattle were higher ($p < 0.05$) than buffalo (Table 3).

Regardless of the type of the animal, TOMD and microbial biomass in SCT silage treated with urea + molasses

Table 3. Gas production parameters of treated sugarcane tops silage by whole rumen microorganisms of cattle and buffalo.

Treatment	Animal	Urea + Molasses (%)	Sulfuric acid (%)	PF* (mg mL ⁻¹)	Microbial biomass (mg)	Efficiency microbial biomass (%)	True organic matter disappearance (mg)	Cell wall degradation
1	Buffalo	0.00	0.00	4.30 ^a	42.00 ^{bc}	49.00 ^a	73.50 ^d	54.83
2		1.00 + 3.00	0.00	3.83 ^b	28.50 ^{dc}	42.00 ^b	67.00 ^c	64.00
3		0.00	2.40	2.58 ^f	25.20 ^d	15.00 ^f	171.50 ^a	65.83
4		1.00 + 3.00	2.40	3.14 ^{de}	50.30 ^{ab}	30.00 ^{de}	168.00 ^a	69.27
1	Cattle	0.00	0.00	3.04 ^e	29.85 ^{dc}	27.00 ^e	107.90 ^b	55.07
2		1.00 + 3.00	0.00	2.92 ^e	24.40 ^d	24.00 ^e	99.20 ^b	61.63
3		0.00	2.40	3.48 ^c	61.30 ^a	37.00 ^{bc}	166.90 ^a	67.50
4		1.00 + 3.00	2.40	3.39 ^{de}	62.75 ^a	35.00 ^{dc}	178.25 ^a	71.03
SEM				0.0866	4.06	0.0192	4.05	-
<i>p</i> -value				0.0001	0.0001	0.0001	0.0001	-
<i>Regardless of the type of the treatment</i>								
Buffalo				3.46 ^a	36.50 ^b	33.97	120.00 ^b	63.48
Cattle				3.21 ^b	44.57 ^a	30.97	138.07 ^a	63.81
SEM				0.0433	2.0300	0.0093	3.1300	-
<i>p</i> -value				0.0001	0.0001	0.0001	0.0001	-
<i>Regardless of the type of the animal</i>								
1		0.00	0.00	3.67 ^a	35.92 ^c	38.21 ^a	90.72 ^b	54.95
2		1.00 + 3.00	0.00	3.37 ^b	26.45 ^b	33.52 ^b	83.10 ^b	62.81
3		0.00	2.40	3.03 ^c	43.25 ^b	25.63 ^b	169.20 ^a	66.66
4		1.00 + 3.00	2.40	3.27 ^b	56.52 ^a	32.52 ^a	173.12 ^a	70.15
SEM				0.0612	2.8700	0.0136	4.4300	-
<i>p</i> -value				0.0001	0.0001	0.0001	0.0001	-

*PF: partitioning factor; SEM: Standard error of the means.

Different superscripts within each column indicate significant differences ($p < 0.05$).

Due to lack of repetition, statistical analysis was not done in terms of cell wall degradation.

+ sulfuric acid (173.12 mg and 56.52 mg, respectively) were higher than other treatments ($p < 0.05$). The highest PF and microbial biomass efficiency were in control (3.67 mg mL⁻¹ and 38.21%, respectively), ($p < 0.05$) (Table 3).

Gas production parameters by rumen bacteria. The highest microbial biomass for buffalo was in SCT ensiled with sulfuric acid (69.10 mg) and in cattle, it was for urea + molasses + sulfuric acid (75.25 mg) treatment. The highest

PF and microbial biomass efficiency for buffalo were in control (9.75 mg mL⁻¹ and 77.42%, respectively) and for cattle, the highest PF and microbial biomass efficiency were in urea + molasses + sulfuric acid treatment (4.37 mg mL⁻¹ and 49.63%, respectively) ($p < 0.05$). In buffalo, TOMD of acid treatment (145.77 mg) and in cattle, TOMD of urea + molasses + sulfuric acid treatment (151.51 mg) had the highest values ($p < 0.05$), (Table 4).

Table 4. Gas production parameters of treated sugarcane tops silage by rumen bacteria of cattle and buffalo.

Treatment	Animal	Urea + Molasses (%)	Sulfuric acid (%)	PF* (mg mL ⁻¹)	Microbial biomass (mg)	Efficiency microbial biomass (%)	True organic matter disappearance (mg)	Cell wall degradation
1	Buffalo	0.00	0.00	9.75 ^a	56.60 ^c	77.42 ^a	62.47 ^c	46.53
2		1.00 + 3.00	0.00	6.57 ^b	37.89 ^d	66.53 ^b	56.95 ^c	50.50
3		0.00	2.40	3.78 ^d	60.89 ^{bc}	41.73 ^d	145.77 ^a	55.88
4		1.00 + 3.00	2.40	4.26 ^c	69.10 ^{ab}	48.34 ^c	142.80 ^a	58.80
1	Cattle	0.00	0.00	4.02 ^{cd}	41.52 ^d	45.20 ^{cd}	91.76 ^b	46.73
2		1.00 + 3.00	0.00	3.08 ^e	24.18 ^e	28.60 ^e	84.32 ^b	52.31
3		0.00	2.40	4.25 ^c	68.52 ^{ab}	48.22 ^c	141.86 ^a	57.30
4		1.00 + 3.00	2.40	4.37 ^c	75.25 ^a	49.63 ^c	151.51 ^a	60.30
SEM				0.1143	3.4500	0.0154	5.3200	-
<i>p</i> -value				0.0001	0.0001	0.0001	0.0001	-
<i>Regardless of the type of the treatment</i>								
Buffalo				6.09 ^a	56.12	58.50 ^a	102.00 ^b	52.92
Cattle				3.93 ^b	52.37	42.91 ^b	117.36 ^a	54.16
SEM				0.0571	1.1300	0.0077	2.6600	-
<i>p</i> -value				0.0001	0.1636	0.0001	0.0001	-
<i>Regardless of the type of the animal</i>								
1		0.00	0.00	6.88 ^a	49.06 ^b	61.31 ^a	77.12 ^b	46.63
2		1.00 + 3.00	0.00	4.83 ^b	31.03 ^c	47.57 ^{bc}	70.63 ^b	51.40
3		0.00	2.40	4.02 ^d	64.71 ^a	44.97 ^c	143.82 ^a	56.59
4		1.00 + 3.00	2.40	4.32 ^c	72.17 ^a	48.10 ^b	147.16 ^a	59.55
SEM				0.0808	2.4400	0.0109	3.7600	-
<i>p</i> -value				0.0001	0.0001	0.0001	0.0001	-

*PF: partitioning factor; SEM: Standard error of the means.

Different superscripts within each column indicate significant differences ($p < 0.05$).

Due to lack of repetition, statistical analysis was not done in terms of cell wall degradation.

Regardless of the type of the treatment, PF, microbial biomass efficiency and microbial biomass by buffalo were more than cattle ($p < 0.05$). The TOMD in cattle was higher than buffalo ($p < 0.05$) (Table 4).

Regardless of the type of the animal, the TOMD and microbial biomass of SCT silage treated with urea + molasses + sulfuric acid ($p < 0.05$) and PF and microbial biomass efficiency in control (6.88 mg mL⁻¹ and 61.30%, respectively), were the highest ($p < 0.05$) (Table 4).

Discussion

Treatments containing sulfuric acid or urea + molasses + sulfuric acid had the highest rate of GP, probably due to presence of rapidly fermentable carbohydrates in molasses, increased ammonia gas production by adding urea,¹⁹ and also neutral detergent fiber (NDF) reduction through degradation of hemicellulose bounds by sulfuric acid.²⁰ Our results were in accordance with previous report suggesting that silages treated with hydro chloric acid

and urea were more degradable than untreated ones.²¹ It has also been shown that hydrochloric acid increases rapid degradable fraction of alfalfa silage. This finding has been contributed to the inhibitory effect of acid on deleterious aerobic fermentation.²¹ These results were in agreement with the results of Khosropour,²² who showed that adding 2.40% sulfuric acid to SCT silage, increases GP rate. Generally, adding urea + molasses + sulfuric acid or sulfuric acid individually, improved nutritional value and utilization of SCT silage by WRM.

Regardless of the type of the treatment (Table 1), potential of GP by WRM of cattle was significantly higher than buffalo, but GP rate was higher in buffalo. The lower GP rate means little GP during the early hours of incubation in cattle, probably due to the fact that fungi have greater role in fiber digestion than other micro-organisms and their colonization is dilatory and needs more time for digestion of feeds and consequent GP.²³ Results of the present experiment are in agreement with, Shakarami,¹² who described that GP rate from wheat

straw by WRM of buffalo is higher than cattle. Our results were in contrast to findings of Rafiee,¹¹ who reported that GP rate of wheat straw by the WRM of buffalo is higher than cattle. The higher potential of GP in cattle compared to buffalo, might be related to higher amounts of fungi in rumen of cattle than buffalo, which have important role in fiber digestion. According to Kumar *et al.* report,²⁴ with diet based on rye grass-concentration, rumen fungi in cattle were higher than buffalo. It has also been reported that for diet based on wheat straw, the rumen fungi of Khouzestan buffalo (2.00×10^3 mL) are lesser than Holstein cattle (2.70×10^3 mL).¹² Whereas Chanthakhoun *et al.* and Wanapat reported that anaerobic fungi of buffalo are more than cattle depending on diet and other conditions.^{25,26} Fungi are about 8.00% of rumen microbial biomass,²⁷ and digest more than 70.00% of cellulose,²⁸ and about 34.00% of plant tissues lignin.²⁹ It has been reported that anaerobic fungi increase feed efficiency and nutrient digestibility of crossbred calves.³⁰ Other reasons for the differences in digestion and its improvement in cattle may be attributed to ability of cattle for digestion of cell wall components. It was found that in fibrous ration, NDF and acid detergent fiber (ADF) digestibility in buffalo is lower than cattle, which could be due to better utilization of cellulose in cattle than buffalo.³¹ During feeding with diet based on sorghum, Moran *et al.* observed same results for Short Horn beef cattle compared to buffalo.³² Adding concentrate to wheat straw based diet, significantly increased digestibility of dry matter (DM), crude protein and NDF in cattle, while in buffalo only increased the digestibility of crude protein.³³ Therefore, in the present experiment, in addition to higher fungi population, higher GP in cattle than buffalo, might be because of more ability of cattle for nutrients digestion, especially fiber.

Regardless of the type of the animal (Table 1), the highest potential and rate of GP were numerically in the treatment containing 2.40% sulfuric acid. The reason of increased potential and rate of GP by adding sulfuric acid was probably due to the effect of chemical treatments such as acid for degradation of lignocellulosic linkages between structural components and increasing their bioavailability for microorganisms.³⁴ Haddi *et al.* reported that there is significant negative correlation between NDF and ADF of feed and the rate and potential of GP.³⁵ The negative effect of cell wall content on GP could be due to reduction of microbial activity via increasing adverse environmental conditions by duration of incubation time. Since SCT contain high amounts of NDF (76.96%) and acid detergent lignin (6.69%),³⁶ the chemical treatment loosens and breakdowns the ester linkages resulting in NDF reduction,³⁷ and digestibility and GP rate increase. On the other hand, urea + molasses has high disappearance rate, but when silage was opened, probably little amount of them remains. Thereby, in comparison to control, this treatment had no significant effect on GP rate.

Addition of urea + molasses + sulfuric acid numerically decreased potential of GP compared to control ($p > 0.05$). Probably this effect was due to high amount of sulfur in medium. By contemporary using of molasses and sulfuric acid (as source containing sulfur), high sulfur resulted in reduction of rumen bacteria growth, especially cellulolytic ones and NDF digestibility. Another reason is probably due to high reduction of rumen pH following simultaneous adding of urea + molasses and sulfuric acid. When rumen pH is lower than 6.20, DM and fiber degradation significantly reduces.³⁸

The treatments had significant effect on GP (Table 2) of SCT silage by rumen bacteria of cattle and buffalo. Treatment with sulfuric acid increased the potential and rate of GP. Literatures showed that acid causes hemicelluloses degradation and NDF reduction leading to GP increase.³⁹ Feeds resources with lower NDF have higher potential of GP. Our results are in agreement with Chaji and mohammadabadi, who reported that treating sugarcane pith by sulfuric acid increases GP by rumen bacteria.⁴⁰ Behgar *et al.* reported that using sulfuric acid + formic acid causes DM degradability increase of alfalfa silage.⁴¹ Generally, the chemical additives such as acid increase potential and effective degradability of material by rumen microorganisms.⁴²

Regardless of the type of the treatment (Table 2), in the present study, the difference between buffalo and cattle rumen bacteria fermentation could be explained by different microbial activity of them. These results are in agreement with those reported by Calabro *et al.* who found that *in vitro* GP of forages feeds using inoculums of buffalo rumen is lower than cattle.⁴³ Paul and Lal reported that GP by inoculums of buffalo rumen is lower than cattle, which is related to less methane production in buffalo.⁴⁴ However, Chanthakhoun *et al.* reported that GP by inoculums of buffalo rumen is more than cattle.⁴⁵ As previously mentioned, the reasons of antithetic results can be related to physiological factors such as age, feeding behavior, level of production, animal health, the nature and relationships between different microbial populations and also external factors such as diet composition, nature of feed, feed frequency, dietary changes, change of seasons, changes in day length and geographical factors, which affect the ratio and density of different groups of rumen microorganisms.¹³

Regardless of the type of the animal (Table 2), adding 2.40% sulfuric acid resulted in the highest potential and rate of GP, probably because of degradation of cellulose and lignin ester barriers and subsequent increased biological usability of microorganisms.⁴⁶ Therefore, the results showed that treatment of SCT with 2.40% sulfuric acid or urea + molasses + acid increases GP, may be because of synchronization between readily fermentable carbohydrates of molasses and urea nitrogen and degradation or loosening of lignocellulose bounds.²¹

The PF and microbial biomass efficiency were the highest in buffalo for control, because due to lack of available nutrients (no additive) in control, GP was very low; since PF is calculated as the ratio of truly degraded substrate to GP volume, it also reflects the variation of short chain fatty acids production per unit of degraded substrate.^{18,46} In cattle, higher PF in treatment containing acid indicated that more organic matter is converted to microbial biomass, so the efficiency of microbial protein synthesis is higher than other treatments. In sulfuric acid treated silages, large amounts of carbohydrates and structural proteins are easily degraded. Therefore, using acid during ensiling, prevents from destroying of proteins, degrades bounds between cellulose, hemicellulose and lignin and subsequently increases the digestibility of DM and crude fiber.⁴⁷ The high part of increasing cell wall degradation is due to degradation of molasses carbohydrates, which increases the growth of cellulolytic bacteria. In addition, increased availability of crude protein by adding urea resulted in increased digestibility. Urea in rumen can convert to ammonia, and ammonia is the sole nitrogen source for most cellulolytic bacteria, so urea + molasses increases the amount of cell wall degradation.⁴⁸

Regardless of the type of the treatment (Table 3), microbial biomass and TOMD of cattle were more than buffalo, probably because of higher ability of cattle in cell wall digestibility or more rumen fungi population of cattle.¹² There is a high negative correlation between structural carbohydrates, particularly lignin, and organic matter (OM) digestibility for both inoculants of cattle and buffalo.⁴³ These observations are in agreement with findings of Bhatia *et al.* who reported that amount of microbial protein synthesis in cattle, based on wheat straw-alfalfa hay-concentrate and wheat straw-clover diets is higher than Indian buffalo.³³ Kennedy *et al.* also reported that with fibrous diets, NDF digestibility in buffalo is lower than cattle, probably due to better utilization of cellulose in cattle than buffalo, which is consistent with our results.³¹

The treatments containing urea + molasses + sulfuric acid and sulfuric acid individually (Table 4) showed the highest GP parameters. Treatment with urea caused ammonia production and due to improving of carbohydrate and nitrogen synchronization, the accessibility of rumen or silage microbes to cell wall polysaccharides increased.⁴⁹ In the rumen, hydrolysis rate of urea is fast, so released ammonia is not utilized efficiently for synthesis of microbial protein. In order to slow down the release of various complexes of urea, using starch and molasses has been recommended.¹⁹ In the rumen, synchronization between nitrogen (e.g. urea) and carbohydrate sources (e.g. molasses) is important.⁵⁰ In the present study, using sulfuric acid in silage prevented from degradation of protein and carbohydrates to non-protein nitrogen and organic acids, respectively,⁴⁷ thus nutrients for bacteria

increased. Salari showed that treatment of palm leaves with urea and molasses increases OM digestibility that is in accordance with the results of the present experiment.⁵¹

Regardless of the type of the treatment (Table 4), PF, microbial biomass efficiency and microbial biomass by buffalo were more than cattle. The TOMD in cattle was higher than buffalo. Similar to the present experiment conditions, Rafiee¹³ reported that PF, microbial biomass and microbial biomass efficiency of wheat straw by buffalo are significantly higher than cattle, may be due to higher bacterial populations of buffalo than cattle. Reportedly, the number of cellulolytic bacteria in swamp buffalo is higher than cattle when both are fed with diet based on rice straw.²⁵ The majority of microbial studies also showed that under the same nutritional conditions, whole bacterial population and cellulolytic, proteolytic, amylolytic and lipolytic bacteria in buffalo are higher than cattle.⁵²

In conclusion, treatments or additives used in the presents study were the proper methods for prolonged preservation of SCT in silo. In aspects of comparison between cattle and buffalo, WRM and bacteria of cattle showed higher digestion of treated SCT silage than buffalo. Addition of sulfuric acid not only had no negative effect on microorganisms, particularly bacteria, but also due to presence of sulfur in sulfuric acid, had positive effect on nutrients digestibility and growth of microorganisms. Thus, using sulfuric acid individually or simultaneously with urea + molasses is the effective and safe method to prepare SCT silage.

References

1. FAO. In: FAO Year Book. Food and Agriculture Organization of United Nations 2012: 181.
2. Ortiz-rubio MA, Ørskov ER, Milne J, et al. Effect of different sources of nitrogen on *in situ* degradability and feed intake of zebu cattle fed sugarcane tops (*Saccharum officinarum*). *Anim Feed Sci Technol* 2007; 139 (3-4): 143-158.
3. Eun JS, Beauchemin KA, Hong SH, et al. Exogenous enzymes added to untreated or ammoniated rice straw: Effects on *in vitro* fermentation characteristics and degradability. *Anim Feed Sci Technol* 2006; 131 (1-2):87-102.
4. Ghoorchi T, Ghanbari F, Ebrahimi T. Evaluation of the effect of adding various additives on aerobic stability, chemical composition and microbes corn silage [Farsi]. *Iranian J Vet Res* 2012; 4 (4): 335-344.
5. Mahmodi-Meymand S, Chaji M, Eslami M, et al. Preparation of sugarcane top silage by different additives and evaluation of its nutritional value by *in vitro* methods [Farsi]. *J Livest Res* 2015; 2 (4): 11-23.
6. Boodoo AA, Delaitre JC, Preston TR. Ensiling sugarcane tops with different additives. *Trop Anim Health Pro* 1977; 2: 2.

7. Baytak E, Aksu T, Muruz H. The effects of formic acid, molasses and inoculants as silage additives on corn silage composition and ruminal fermentation characteristics in sheep. *J Vet Anim Sci* 2005; 29 (2): 469-474.
8. Slottner D, Bertilsson J. Effect of ensiling technology on protein degradation during ensilage. *Anim Feed Sci Technol* 2006; 127 (1-2): 101-111.
9. Promkot C, Wanapat M. Effect of sulfur on rumen ecology, blood metabolites, hormones and production in lactating dairy cows supplemented with fresh cassava foliage or cassava hay. In proceedings: Mekarn regional conference: Matching livestock systems with available resources. Halong Bay, Vietnam. 2007: 25-28.
10. Animal husbandry and veterinary medicine. Effect of inorganic sulfur on the amount of microbe in rumen and the metabolism of nitrogen and sulfur in sheep. Available at: www.agrpaper.com. Accessed: May 28, 2012.
11. Rafei M, Chaji M, Mohammadabadi T, et al. The comparison of digestibility of wheat straw by rumen bacteria and rumen microorganisms of Holstein cow and Khuzestan water buffalo [Farsi]. *J Anim Sci Res* 2015; 25 (2): 43-55.
12. Shakarami F, Chaji M, Eslami M, et al. The comparison of *in vitro* digestibility of wheat straw by rumen anaerobic fungi of Khuzestan buffalo and Holstein cattle. *Iranian J Appl Anim Res* 2015; 5(2): 285-292.
13. Russell JB, Strovel HJ, Chen GJ. Enrichment and isolation of a ruminal bacterium with a very high specific activity of ammonia production. *Appl Environ Microb* 1988; 54 (4): 872-877.
14. Menke KH, Steingass H. Estimation of the energetic feed value obtained from chemical analysis and *in vitro* gas production using rumen fluid. *Anim Res Dev* 1988; 28: 7-55.
15. Mohammadabadi T, Chaji M, Bojarpour M. The Effect of processing of sugarcane pith with steam on gas production parameters by using isolated rumen microbiota [Farsi]. *Iranian Res J Anim Sci* 2012; 4(3): 240-246.
16. Ørskov ER, McDonald I. The estimation of protein degradability in the rumen from incubation measurements weighted according to rate of passage. *J Agr Sci* 1979; 92 (2): 499-503.
17. Olivera MP. Use of *in vitro* gas production technique to assess the contribution of both soluble and insoluble fraction on the nutritive value of forage. MSc Thesis. University of Aberdeen, Scotland. 1998.
18. Makkar HPS. *In vitro* gas methods for evaluation of feeds containing phytochemicals. *Anim Feed Sci Technol* 2005; 123-124: 291-302.
19. Nisha J, Tiwari S, Singh P. Effect of urea molasses mineral granules (UMMG) on rumen fermentation pattern and blood biochemical constituents in goat kids fed sola (*Aeschynomene indica Linn*) grass-based diet. *Vet Arch* 2005; 75(6): 521-530.
20. Valizadeh R, Naserian AA, Ajdarifard A. Silage biochemistry [Farsi]. Mashhad, Iran: Ferdowsi University of Mashhad Press 2001: 405.
21. Vakili AR, Daneshmegharan M, Moghaddam HN. Specific chemical, degradability dry matter parameters and crude protein alfalfa silage richest with HCL and urea and its effect on production characteristic of early lactating Holstein cows [Farsi]. *Iranian J Anim Sci* 2009; 1(19): 54-62.
22. Khosropour V. The investigation of the effect processing of sugarcane top by chemical and enzymatic methods on improvement nutritional value in ruminant. MSc Thesis. Department of Animal Science, Ramin Agriculture and Natural Resources University of Khuzestan, Iran 2012.
23. Sangwan DC, Kumar S, Bhatia SK, et al. Comparative *in vitro* rumen fungal degradation and gas production of cellulosic feeds in cattle and buffalo. *Indian Coun Agr Res* 2002; 72 (2): 174-179.
24. Kumar S, Sing S, Bhatia SK. Microbial and biochemical change in the rumen of cattle and buffalo fed oat hay concentrate diet. *Indian J Anim Nutr* 2002; 19: 78-79.
25. Chanthakhoun V, Wanapat M, Kongmun P, et al. Comparison of ruminal fermentation characteristics and microbial population in swamp buffalo and cattle. *Livest Sci* 2012; 143(2-3): 172-176.
26. Wanapat M. Potential uses of local feed resources for ruminants. *Tropic Anim Health Pro* 2009; 41(7): 1035-1049.
27. Bauchop T. Rumen anaerobic fungi of cattle and sheep. *Appl Environ Microb* 1979; 38(1): 148-158.
28. Akin DE, Borneman WS. Role of rumen fungi in fiber degradation. *J Dairy Sci* 1990; 73(10): 3023-3032.
29. Krause DO, Denman SE, Mackie RI, et al. Opportunities to improve fiber degradation in the rumen: Microbiology, ecology and genomics. *FEMS Microb Rev* 2003; 27(5): 663-693.
30. Dey A, Sehgal JP, Puniya AK, et al. Influence of anaerobic fungal culture (*Orpinomyces sp.*) administration on growth rate, ruminal fermentation and nutrient digestion in calves. *Asian Australas J Anim Sci* 2004; 17(6): 820-824.
31. Kennedy PM, McSweeney CS, Foulkes D, et al. Intake and digestion in swamp buffaloes and cattle. 1. The digestion of rice straw (*Oryza sativa*). *J Agr Sci* 1992; 119(2): 227-242.
32. Moran JB, Norton BW, Nolan JV. The intake, digestibility and utilization of a low-quality roughage by Brahman cross, buffalo Banteng and Shorthorn steers. *Aust J Agr Res* 1979; 30(2): 333-340.
33. Bhatia SK, Kumar S, Sangwan DC. Advances in buffalo-cattle nutrition and rumen ecosystem. Locknow, India: International Book Distributing Co. 2004: 163.
34. Rowghani E, Zamiri MJ. The effects of a microbial inoculant and formic acid as silage additives on

- chemical composition, ruminal degradability and nutrient digestibility of corn silage in sheep. *Iranian J Vet Res* 2009; 10 (2): 110-111.
35. Haddi ML, Filacorda S, Meniai, et al. *In vitro* fermentation kinetics of some halophyte shrubs sampled at three stages of maturity. *Anim Feed Sci Technol* 2003; 104(1-4): 215-225.
36. Gendley MK, Singh P, Garg AK. Performance of crossbred cattle fed chopped green sugarcane tops and supplemented with wheat bran or lentil chuni concentrates. *Asian Australas J Anim Sci* 2002; 15(10): 1422-1427.
37. Liu JX, Ørskov ER. Cellulase treatment of untreated and steam pre-treated rice straw effect on *in vitro* fermentation characteristic. *Anim Feed Sci Technol* 2000; 88(3-4): 189-200.
38. Cronje PB. Ruminant physiology: Digestion, metabolism, growth, and reproduction. Peretoria, South Africa: CABI Publishing 2000; 282-305.
39. Sommart K, Parker DS, Rowlinson, et al. Fermentation characteristics and microbial protein synthesis in an *in vitro* system using cassava, rice straw and dried ruzi grass as substrates. *Asian Australas J anim Sci* 2000; 13(8): 1084-1093.
40. Chaji M, Mohammadabadi T. The effect of low temperature steam, sodium hydroxide and exogenous enzyme on *in vitro* degradation of rice straw by rumen bacteria of sheep. *Indian J Small Rumin* 2010(2); 190-194.
41. Behgar M, Daneshmesgaran M, Nasiri Moghadam H, et al. Chemical composition, dry matter degradability and crude protein of alfalfa silage the processing with formic and sulfuric acids and it effect on Holstein cows [Farsi]. *J Sci Natur Res* 2007; 40 (11): 339-349.
42. Castro FB. The use of steam treatment to upgrade lingo cellulosic materials for animal feed. PhD. Thesis. University of Aberdeen, UK; 1994.
43. Calabro S, Moiello G, Picclo V, et al. Rumen fermentation and degradability in buffalo and cattle using the *in vitro* gas production technique. *J Anim Physio Anim Nutr* 2008; 92(3): 356-362.
44. Paul SS, Lal D. Nutrient Requirements of Buffaloes. Dehli, India: Satish Serial Publishing House 2010; 5-17.
45. Chanthakhoun V, Wanapat M, Wanapat S. Various feeds using rumen fluid from swamp buffalo and beef cattle by *in vitro* gas fermentation technique. In proceedings: Agriculture conference. Khon Kaen, Thailand. 2009; 25-27.
46. Blummel M, Makkar HPS, Becker K. *In vitro* gas production: A technique revisited. *J Anim Physio Anim Nutr* 1997; 77(1-5): 24-34.
47. McDonald P, Henderson AR, Heron SJE. The biochemistry of silage. 2nd ed. Marlow, UK: Chalcombe Publications 1997: 83.
48. Alikhani M, Asadi A, Ghorbani G, et al. Chemical composition and dry matter degradability of sunflower silage as influenced by addition of urea, molasses and inoculants [Farsi]. *Iranian J Sci Technol Agr and Natur Res* 2005; 9 (3): 171-182.
49. Horn HW, Zarrila-Rios J, Akin DE. Influence of stage of forage maturity and ammoniation of wheat straw on ruminal degradation of wheat forage tissue. *J Anim Sci Tech* 1989; 24(3-4): 201-218.
50. Choi CW, Vanhatalo A, Ahvenjaru S, et al. Effect of several protein supplement on flow of soluble non-ammonia nitrogen from the forestomach and milk production in dairy cows. *Anim Feed Sci Technol* 2002; 102(1-4): 15-33.
51. Salari Sarduei B. The using of date leaf in nutrition productive Raeeni goats. In proceedings: The 4th congress on animal science. Karaj, Iran. 2010; 1-5.
52. Singh S, Pradhan K, Bhatia SK, et al. Relative rumen microbial profile of cattle and buffalo fed wheat straw-concentrate diets. *Indian J Anim Sci* 1992; 62: 1197.