

Effect of salinity level on TSH and thyroid hormones of grass carp, *Ctenophayngodon idella*

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Article Info	Abstract
<p>Article history:</p> <p>Received: 25 August 2012 Accepted: 08 December 2012 Available online: 15 September 2013</p> <p>Key words:</p> <p>Grass carp Salinity Serum Thyroid hormones Thyrotropin</p>	<p>Thyroid hormones (T₃, T₄) have marked effect on body metabolism and in controlling osmoregulation activity in fish. The aim of this study was to determine the effect of water salinity changes on thyroid hormones level and thyroid-stimulating hormone (TSH) of grass carp. For this purpose 120 grass carp were divided randomly in to four groups (10 fish in each group and three replicates per treatment). Three groups were held in three different salinities at concentrations of 4, 8 and 12 g L⁻¹. The fourth group was reared in fresh water and considered as control. After three weeks blood samples were collected from the caudal peduncle vein. Then serum was separated and serum thyroid hormones and TSH were measured by LISA on Microwell plates. Our results indicated that the average of T₃ levels in 4, 8 and 12 g L⁻¹ groups were 0.43 ± 0.11, 0.22 ± 0.04 and 0.21 ± 0.04 µg dL⁻¹, respectively. T₃ levels in all experimental groups were significantly lower than those of control group ($p < 0.05$). Serum T₄ level in 4, 8 and 12 g L⁻¹ groups were 0.29 ± 0.06, 0.24 ± 0.05 and 2.85 ± 0.55 µg dL⁻¹, respectively. Thyroxine level was significantly higher only in 12 g L⁻¹ group in comparison with the control and other experimental groups ($p < 0.05$). Thyroxine level in other groups had not any significant difference with the control group ($p > 0.05$). The level of TSH in salinities of 4 and 8 g L⁻¹ groups was significantly higher than that of control group ($p < 0.05$). The results showed that increasing water salinity can have significant effect on thyroid activity by decreasing T₃ and increasing T₄ level in serum of grass carp in experimental condition.</p> <p>© 2013 Urmia University. All rights reserved.</p>

بررسی تأثیر سطوح مختلف شوری بر TSH و هورمون‌های تیروئیدی سرم خون ماهی کپور علفخوار *Ctenophayngodon idella*

چکیده

هورمون‌های تیروئیدی (T₃، T₄) تأثیرات مهمی بر روی متابولیسم بدن و کنترل فعالیت تنظیم‌اسمزی در ماهی دارند. هدف از انجام این مطالعه مشخص کردن تأثیر تغییرات شوری آب بر سطح هورمون‌های تیروئیدی و هورمون محرک تیروئید (TSH) ماهی کپور علفخوار بوده است. برای اینکار ۱۲۰ قطعه ماهی کپور علفخوار در معرض سه غلظت مختلف نمک دریایی قرار داده شدند و یک گروه نیز به عنوان شاهد در نظر گرفته شد. پس از گذشت سه هفته خونگیری و جداسازی سرم انجام شد و میزان هورمون‌های تیروئیدی و TSH با روش الیزا اندازه‌گیری شد. در این مطالعه میانگین سطح سرمی T₃ در گروه‌های شوری ۴، ۸ و ۱۲ گرم در لیتر به ترتیب ۰/۱۱ ± ۰/۴۳، ۰/۰۴ ± ۰/۲۲ و ۰/۰۴ ± ۰/۲۱ میکروگرم در دسی لیتر بوده است که این میزان‌ها نسبت به گروه شاهد کاهش معنی‌داری داشته است ($p < 0.05$). میانگین سطح سرمی T₄ در گروه‌های شوری ۴، ۸ و ۱۲ گرم در لیتر به ترتیب ۰/۰۶ ± ۰/۲۹، ۰/۰۵ ± ۰/۲۴ و ۰/۵۵ ± ۲/۸۵ میکروگرم در دسی لیتر بوده است که تنها در گروه شوری ۱۲ گرم در لیتر در مقایسه با گروه شاهد و دیگر گروه‌ها افزایش معنی‌داری داشته است ($p < 0.05$) و در گروه‌های دیگر اختلاف معنی‌داری با گروه شاهد نداشته است ($p > 0.05$). میانگین سطح سرمی TSH در شوری‌های ۴ و ۸ گرم در لیتر به طور معنی‌داری بالاتر از گروه شاهد بوده است ($p < 0.05$). نتایج نشان داد که افزایش شوری آب می‌تواند باعث تأثیر بر فعالیت تیروئید شده و موجب کاهش معنی‌دار هورمون T₃ و افزایش معنی‌دار هورمون T₄ در سرم ماهی کپور علفخوار شود.

واژه‌های کلیدی: تیروتروپین، سرم، شوری، ماهی کپور علفخوار، هورمون‌های تیروئیدی

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Introduction

Salinity stress is one of the most common stresses in freshwater aquatic animals that usually occurs and if prolonged, can lead to reduction of production efficiency or fish death. Carps and other bonny fishes are able to regulate the ionic concentration of the internal body fluids within narrow ranges. In many studies, the exposure of freshwater fish to high salinities (salinity stress), is able to change the internal osmolarity of the plasma allowing an influx of water.¹⁻⁴ Different hormones control the activities of the organs involved in osmoregulation, which maintain water and mineral balance in different salinity.⁵ Thus, conditions that affect or disrupt the ionic homeostasis, cause changes in hormone levels involved in osmoregulation as a response to control the ion transport capacity.^{6,8} The thyroid hormones, thyroxine (T_4) and tri-iodothyronine (T_3) are products of the thyroid gland in all vertebrates. Thyroid stimulating hormone (Thyrotropin, also known as TSH), T_4 and T_3 are the principal hormones secreted from the hypothalamic-pituitary-thyroid (HPT) axis; produce a wide range of physiologic actions in fish. Thyroid hormones of fish are small molecules very similar to other vertebrates. They are called T_4 and T_3 in fish. Tri-iodothyronine is the most effective biological thyroid hormone in fish.⁹ Thyroid hormones are important to control development, growth, metabolism and osmoregulation in fish and often do these activities in association with other hormones such as cortisol and growth hormone.¹⁰ Grass carp as a freshwater fish may be exposed to salinities higher than normal range. In some parts of Khuzestan province water salinity of fish ponds may be higher than usual in some months of the year. Since there are few studies on the thyroid hormones of grass carp, it was necessary to study the normal level as well as the effect of increase salinity on thyroid hormones level in grass carp.

Materials and Methods

Experimental groups. In this study 120 grass carp (100 to 120 g weight range) were divided randomly into four groups (10 fish in each group and three replicates per treatment). Three groups were held in three different salinities at concentrations of 4, 8 and 12 g L⁻¹. The fourth group was reared in freshwater (1 g L⁻¹) and considered as control. The gradual increase of salinity in experimental groups was done in 3-4 days. All other physicochemical parameters of water and feeding level were identical for all groups.

Sampling and measurements. After three weeks, the fish were anesthetized with 100 mg L⁻¹ MS-222 (Sigma-Aldrich, St. Louis, MO, USA) by short term bath method and blood samples were collected from all fishes. For blood collecting, bleeding was done from the caudal peduncle vein. Blood samples were immediately transferred to sterile

tubes and the serum was separated by centrifugation. In this study, T_3 , T_4 and TSH were measured by enzyme linked immunosorbent assay (ELISA) method. Serum free T_4 (f T_4) level was measured manually by ELISA using commercially available kit (Delaware Biotech Inc., Heidelberg, Germany) according to the manufacturer's instructions. For total T_3 and TSH concentration in serum, immune enzymatic colorimetric method was used for quantitative determination using DiaMetra kit (DiaMetra Co., Milano, Italy) according to the manufacturer's instructions. Absorbance was read at 450 nm in ELISA reader (Synergy HT, Winooski, VT, USA).

Statistical analysis. Means of each parameter and changes were measured and compared in all groups by one way analysis of variance. The data were analyzed by SPSS (Version 16, SPSS Inc., Chicago, IL, USA) and the differences were considered significant at level of $p < 0.05$.

Results

According to our results, the average of serum T_4 , T_3 and TSH in control and 4, 8 and 12 g L⁻¹ salinity groups were showed in Table 1. The level of T_4 was significantly higher only in 12 g L⁻¹ group in comparison with control and the other experimental groups ($p < 0.05$). The level of T_4 in other groups did not have any significant difference with control group ($p > 0.05$). The level of T_3 in all experimental groups were significantly lower than the control group ($p < 0.05$). The level of TSH in 4 and 8 g L⁻¹ salinity groups was significantly higher than control group ($p < 0.05$).

Table 1. The average of serum thyroid hormones in grass carp in experimental groups and the control (Mean \pm SE).

Group	T_4 ($\mu\text{g dL}^{-1}$)	T_3 ($\mu\text{g dL}^{-1}$)	TSH ($\mu\text{g dL}^{-1}$)
Control	0.91 \pm 0.19 ^a	1.62 \pm 0.23 ^a	0.13 \pm 0.01 ^a
4 (g L ⁻¹)	0.29 \pm 0.06 ^a	0.43 \pm 0.10 ^b	0.32 \pm 0.05 ^b
8 (g L ⁻¹)	0.24 \pm 0.05 ^a	0.22 \pm 0.04 ^b	0.30 \pm 0.04 ^b
12 (g L ⁻¹)	2.85 \pm 0.55 ^b	0.21 \pm 0.04 ^b	0.25 \pm 0.02 ^a

^{ab} Different letter in each column show significant difference between each hormone concentration ($p < 0.05$).

Discussion

Salinity changes can be stressful and may affect hemostasis of the fish. Numerous experiments on animals demonstrated that blood concentrations of thyroid hormones are increased during early exposure to stressors such as high ambient temperatures and excitement stress.¹¹ Environmental factors such as day length, lunar cycle events, temperature, pH, salinity, and nutrition are all implicated in stimulating thyroid activity.¹² In some fish species, thyroid hormones support the action of growth hormone and cortisol in promoting seawater acclimation.¹³ In our study we observed that with increasing salinity at concentrations of 4, 8 and 12 g L⁻¹, serum T_3 level in these three groups was decreased significantly in comparison with the control group. This decrease in T_3 could be due to

decreased hormone decomposition and delay or decrease of hormone secretion.¹¹

Serum T₄ level in the group with 12 g L⁻¹ salinity was increased significantly in comparison with other groups. The level of TSH in the groups with 4 and 8 grams per liter salinity were significantly higher than that of control group. Parker and Specker investigated the effect of salinity and temperature on whole-animal thyroid hormone levels in larval and juvenile striped bass, *Morone saxatilis*. Their results revealed that higher salinities increased T₄ levels in fish larvae. They suggest that thyroid hormones may mediate the beneficial effects of salinity on larval striped bass growth and survival.¹⁴ Sampaio and Bianchini investigated salinity effects on osmoregulation and growth of the euryhaline flounder *Paralichthys orbignyanus*, and suggest that the lower growth rate exhibited by *P. orbignyanus* in fresh water could be due, at least partially, to a higher energy expenditure associated to a higher branchial Na⁺, K⁺-ATPase activity in this environment.³

Most studies that involve exposing anadromous salmonid fishes to seawater report an increase in gill Na⁺ K⁺-ATPase activity suggesting that it is an integral part of their successful acclimation.¹⁵ Rejitha *et al.* investigated short-term salinity acclimation demands of thyroid hormone action in the climbing perch *Anabas testudineus* Bloch. Their results suggest that following transfer of fish to 20 ppt salinity for a day after transient salinity changes, plasma T₄ was elevated, and plasma T₃ decreased whereas plasma cortisol remained unchanged. The levels of these hormones, however, returned to basal levels when these fish were kept for a prolonged acclimation of three weeks. Their results indicate that salinity acclimation in climbing perch demands thyroid hormone secretion and its action and not cortisol as part of coordinating the acclimation processes in the early phase of salinity acclimation.¹⁶ In other experiment *Acanthopagrus latus* was held in different salinities and results indicated that thyroid hormone levels in plasma (T₃) 12 hr after the salinity change compared to the control samples showed a small rise that quickly return to normal.¹⁷ Saunders *et al.* studied the effect of orally administered 3, 5, 3'-triiodo-L-thyronine on growth and salinity tolerance of Atlantic salmon (*Salmo salar*). They found that T₃ treatment significantly increased salinity tolerance in mature males but not in immature fish.¹⁸ Klaren *et al.* investigated the effect of acclimation to low salinity water of gilthead seabream (*Sparus auratus*) on the activities of thyroid hormone-metabolizing enzymes in gills, kidney, and liver. The data reveal that following acclimation to low salinity water, the plasma free T₄ concentration increases 2.5 fold, and outer ring deiodination activities towards T₄, 3,5,3'-tri-iodothyronine and 3,3',5'-tri-iodothyronine (reverse T₃ also known as rT₃) in the gills are reduced by 20-32%.

Their results substantiate that thyroid hormones are involved in *S. auratus* osmoregulation, and that the gills are

well equipped to play an important role in the modulation of plasma hormone titers.¹⁹ McCormick and Saunders showed that in juvenile Atlantic salmon (*Salmo salar*) following acute exposure to seawater (30 ppt), plasma T₄ was increased 80% in the first 6 hr, declined to initial levels after 24 hr, and remained stable for 18 days, thereafter. In non-smolts, plasma T₄ did not rise immediately after exposure to seawater, fell slightly after 2 days, and remained low for 18 days. Plasma T₃ of smolts and non-smolts was not affected by acute exposure to seawater.²⁰ In other experiment *Solea senegalensis* was acclimated to seawater (38 ppt) and that were transferred and allowed to acclimate to different salinities (5, 15, 38 and 55 ppt) for 17 days. Plasma cortisol had increased, whereas plasma fT₄ levels were decreased in animals that were transferred to salinities other than SW. The inverse relationship between plasma cortisol and fT₄ suggests an interaction between these hormones during osmoregulatory period.²¹ Iwata *et al.* studied the effects of T₄, growth hormone and cortisol on salinity preference of juvenile Coho salmon (*Oncorhynchus kisutch*). Fish treated with T₄ or with ovine growth hormone showed a significantly greater preference for 14 ppt salinity than the controls. Their finding that thyroid hormone, growth hormone and cortisol all stimulate salinity preference of young Coho salmon, suggested the importance of these hormones in osmoregulation and preparing smolts for migration.²²

In conclusion according to our results, the thyroid hormones changes in salinity stress in grass carp showed that these hormones may be involved in maintenance of hemostasis in this fish species after salinity changes. Tri-iodothyronine reduction if remain low, may affect body growth or even death of the fish in prolonged exposure to high salinities. Therefore, in this situation parenteral and/or oral administration of T₃ may increase salinity tolerance. However, for this suggestion it needs more investigation in different ages and conditions.

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References

1. Morgan JD, Iwama GK. Effects of salinity on growth, metabolism, and ion regulation in juvenile rainbow and steelhead trout (*Oncorhynchus mykiss*) and fall Chinook salmon (*Oncorhynchus tshawytscha*). *Can J Fish Aquat Sci* 1991; 48(11): 2083-2094.
2. Van der Linden A, Vanaudenhove M, Verhoye M, et al. Osmoregulation of the common carp (*Cyprinus carpio*) when exposed to an osmotic challenge assessed in-vivo and non-invasively by diffusion- and T2-weighted magnetic resonance imaging. *Comp Biochem Phys A* 1999; 124: 343-352.

3. Sampaio LA, Bianchini A. Salinity effects on osmoregulation and growth of the euryhaline flounder *Paralichthys orbignyanus*. *J Exp Mar Biol Ecol* 2002; 269: 187-196.
4. McCormick SD, Sundell K, Björnsson BT, et al. Influence of salinity on the localization of Na⁺/K⁺-ATPase, Na⁺/K⁺/2Cl⁻ cotransporter (NKCC) and CFTR anion channel in chloride cells of the Hawaiian goby (*Stenogobius hawaiiensis*). *J Exp Biol* 2003; 206: 4575-4583.
5. Lee KM, Kaneko T, Katoh F, et al. Prolactin gene expression and gill chloride cell activity in Fugu, *Takifugu rubripes* exposed to a hypoosmotic environment. *Gen Comp Endocrinol* 2006; 149: 285-293.
6. Kelly SP, Woo NYS. The response of sea bream following abrupt hyposmotic exposure. *J Fish Biol* 1999; 55: 732-750.
7. Chang IC, Lee TH, Yang CH, et al. Morphology and function of gill mitochondria-rich cells in fish acclimated to different environments. *Physiol Biochem Zool* 2001; 74: 111-119.
8. Moron SE, Oba ET, De Andrade CA, et al. Chloride cell responses to ion challenge in two tropical freshwater fish, the *Erythrinids Hoplias malabaricus* and *Hoplerythrinus unitaeniatus*. *J Exp Zool* 2003; 298: 93-104.
9. Baldisserotto B, Mancera JM, Kapoor BG. Fish osmoregulation. Enfield, NH, USA: Science Publishers 2007; 527.
10. Evans DH. The physiology of fishes. 2nd ed. Boca Raton, FL, USA: CRC Press 1997; 441-463.
11. Guerrini VH, Bertchinger H. Effect of exposure to a hot-humid and hot dry environment on thyroid hormone values in sheep. *Brit Vet J* 1983; 139: 119-128.
12. Grau EG. Environmental influences on thyroid function in teleost fish. *Amer Zool* 1988; 23: 329-335.
13. McCormick SD. Endocrine control of osmoregulation in teleost fish. *Integr Comp Biol* 2001; 41(4): 781-794.
14. Parker SJ, Specker JL. Salinity and temperature effects on whole-animal thyroid hormone levels in larval and juvenile striped bass, *Morone saxatilis*. *Fish Physiol Biochem* 1990; 8(6): 507-514.
15. Bystriansky JS, Richards JG, Schulte PM, et al. Reciprocal expression of gill Na⁺/K⁺-ATPase α -subunit isoforms α 1a and α 1b during seawater acclimation of three salmonid fishes that vary in their salinity tolerance. *J Exp Biol* 2006; 209: 1848-1858.
16. Rejitha V, Peter VS, Subhash MC. Short-term salinity acclimation demands thyroid hormone action in the climbing perch *Anabas Testudineus* Bloch. *J Endocr Repro* 13. 2009; 2: 63-72.
17. Movahedinia A, Savari A, Morovvati H, et al. Endocrine responses of Yellowfin seabream (*Acanthopagrus latus*) in adaptation to different environmental salinities. *J Mar Sci Tech* 2010; 8(3): 1-14.
18. Saunders RL, McCormick SD, Henderson EB, et al. The effect of orally administered 3,5,3'-tri-iodo-L-thyronine on growth and salinity tolerance of Atlantic salmon (*Salmo salar* L.). *Aquacult* 2003; 45(1-4): 143-156.
19. Klaren Peter HM, Guzmán José M, Reutelingsperger SJ, et al. Low salinity acclimation and thyroid hormone metabolizing enzymes in gilthead seabream (*Sparus auratus*). *Gen Comp Endocrin* 2007; 152 (2-3): 215-222.
20. McCormick SD, Saunders RL. Influence of ration level and salinity on circulating thyroid hormones in juvenile Atlantic salmon (*Salmo salar*). *Gen Comp Endocrin* 1990; 78(2): 224-230.
21. Arjona FJ, Vargas-Chacoff L, Martín del Río MP, et al. The involvement of thyroid hormones and cortisol in the osmotic acclimation of Solea senegalensis. *Gen Comp Endocrin* 2008; 155(3): 796-803.
22. Iwata M, Yamauchi K, Nishioka RS, et al. Effects of thyroxine, growth hormone and cortisol on salinity preference of juvenile Coho salmon (*Oncorhynchus kisutch*). *Mar Freshw Behav Phy* 1990; 17(4): 191-201.