Veterinary Research Forum

Journal Homepage: vrf.iranjournals.ir

Seroprevalence of Bluetongue in sheep in Kohgiluyeh and Boyer-Ahmad province, Iran

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Article history:

Received: 05 November 2012 Accepted: 10 March 2013 Available online: 15 December 2014

Key words:

Article Info

Bluetongue ELISA Iran Sheep

Abstract

Bluetongue (BT) is a viral disease of ruminants transmitted by Culicoides biting midges and has the ability to spread rapidly over large distances. The disease occurs almost worldwide between latitudes approximately 35° S and 50° N. Among the numerous diseases of ruminants, BT has gained considerable importance in recent years as one of the best examples of the effects of climate change on disease spread. Sheep are major livestock species in Iran, but studies of BT have not gained the priority compared to other diseases. Thus, the objective of this study was to describe the distribution and seroprevalence of bluetongue virus (BTV) infections in sheep in Kohgiluyeh and Boyer-Ahmad province of Iran, and to identify factors associated with the exposure of these sheep to BTV infection. Sera from 262 apparently healthy sheep were collected during the year 2011. The collected sera of the animals were screened with competitive enzyme like immunosorbent assay (c-ELISA). Two hundred and three (77.48%) out of 262 sera tested were positive to BTV antibodies. Statistically significant differences were found in the seroprevalence BT, between sex and age of sheep (p < 0.001). No statistically significant differences were observed in BTV seroprevalence among different seasons, nor among recently aborted and normally delivered.

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شیوع سرمی زبان آبی در گوسفندان استان کهگیلویه و بویراحمد

چکیده

زبان آبی از بیماری های ویروسی نشخوارکنندگان بوده که به وسیله ی نیش پشه کولیکوئیدس منتقل می گردد و قدرت پخش سریع در فواصل طولانی را دارا می باشد. از لحاظ جغرافیایی، بیماری زبان آبی بین عرض جغرافیایی ۳۵ درجه جنوبی و ۵۰ درجه شمالی در دنیا گسترده شده و یکی از بیماری هایی است که تغییرات آب و هوایی بر میزان شیوع آن در مناطق مختلف تأثیر دارد. در سال های اخیر، توجه به بیماری زبان آبی در ایران، این مطالعه به بررسی سال های اخیر، توجه به بیماری زبان آبی در میان سایر بیماری های ویروسی نشخوارکنندگان افزایش یافته اما با توجه به اندک بودن مطالعات بر روی ویروس زبان آبی در ایران، این مطالعه به بررسی شیوع آلودگی و شناسایی فاکتورهای دموگرافیکی مربوط به این ویروس در گوسفندان استان کهگیلویه و بویراحمد پرداخته است. در این مطالعه ۲۶۲ نمونه سرمی از گوسفندان به ظاهر سالم در طی سال ۱۳۹۰ جمع آوری شد. از روش الایزای رقابتی به منظور بررسی حضور آنتی بادی در سرم گوسفندان استفاده گردید. دویست و سه نمونه (۲۰/۱۴) اما در بین فصل های مختلف و میش هایی که اخیراً سقط داشته بادی ویروس زبان آبی مثبت بودند. اختلافات آماری معناداری در شیوع زبان آبی بین جنس و سن گوسفندان مشاهده شد (۲۰۰۱) اما در بین فصل های مختلف و میش هایی که اخیراً سقط داشته یا زیامان آنها طبیعی بوده، اختلاف معناداری مشاهده نگردید.

واژه های کلیدی: الایزا، ایران، زبان آبی، گوسفند

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Introduction

Bluetongue (BT) is an infectious disease of wild and domestic ruminants caused by bluetongue virus (BTV). The RNA virus belongs to the Orbivirus genus in the Reoviridae family. It is transmitted by biting midges of the genus Culicoides. Up-to now 24 distinct serotypes of the virus have been described. The disease was first described in an imported Merino sheep in South Africa in the 19th century. In 1902 the disease was mentioned as "a malarial catarrhal fever of sheep", and was named as "bluetongue" in 1905. Although BTV may infect many species of ruminants, sheep are usually the most severely affected animals. Viremia in sheep and goats commence from three days post infection and may last up to 54 days. Severe disease occurs most commonly in certain breeds of sheep, but the severity of BT is highly variable.

The virus causes infection and clinical disease in sheep, the primary clinical signs of BTV infection is hemorrhage and ulceration of the mucous membranes in the upper portion of the gastrointestinal tract, including the oral cavity and esophagus. Other signs such as coronitis, laminitis, facial edema, and transient infertility are also seen in sheep. Cattle rarely demonstrate clinical disease.⁶ Clinical signs may be acute and mortality can be up to 70.00% in some flocks of sheep.⁷

The historically the geographical distribution of the BT has been approximately between latitudes of 50° N and 35° S, in temperate and tropical regions of the world.⁸ This area coincides with the distribution of specific species of Culicoides midges that are biological vectors of the virus.^{9,10} Although the relationship between the virus and vector is not yet fully understood, environmental and genetic factors are important determinants of bluetongue activity within the vector and its ecosystem.¹¹

While sheep are a major livestock species in Iran, studies of BT have not been given the same priority as some other diseases. Thus, the objective of this study was to describe the distribution and seroprevalence of BTV infection in sheep in Kohgiluyeh and Boyer-Ahmad province in Iran, in 2011. This province is 15504 Km² and is located between latitude 30°30′ to 31°30′ N and longitude 51° to 52° E in the southwest of Iran (Fig. 1).



Fig. 1. Kohgiluyeh and Boyer-Ahmad province in of Iran.

Materials and Methods

Sampling. A total number of 262 serum samples were collected from apparently healthy sheep of various ages and either sexes during four seasons in the year 2011, from 20 flocks in 17 different locations in Kohgiluyeh and Boyer-Ahmad province. Age was determined by tooth replacement in sheep.¹² The animals were divided into three age groups: juvenile (≤ 1 year old), sub-adult (1 to 3 years old) and adult (> 3 years old). The blood samples were collected from the jugular vein with sterile tubes of venoject (Zhejiang U-REAL medical technology Co., Taizhou, China) without anticoagulant and the sample shipped from zones of sampling to laboratory in dry ice and then centrifuged at 3000 rpm for 15 min. Then, serum was separated and stored at −20 °C until enzyme like immunosorbent assay (ELISA) was performed.

Serological test. The competitive ELISA (c-ELISA) has proved to be the best serologic test for detecting group antibodies to BTV.¹³ In this study, BTV antibody levels were measured using c-ELISA IDEXX bluetongue competition® assay (IDEXX BT, Hoofddorp, The Netherlands) according to the manufacturer's instructions. The optic density of each sample was read by an ELISA microplate reader (PowerWave XS2; BioTek, Vermont, USA) at 450 nm.

Results are expressed as percentage of negativity (PN) compared to the kit control and designated as positive, doubtful or negative according to the cut-off values recommended by the manufacturer (PN \leq 70 is positive; 70 < PN < 80 is doubtful; $PN \geq 80$ is negative). Statistical analyses were performed using a threshold value of 70 that discriminate between positive (PN \leq 70) and negative (PN \leq 70) BTV c-ELISA results.

Statistical analysis. Statistical analyses were performed using SPSS (Version 16.0; SPSS Inc., Chicago, USA). The association between age (categorical; juvenile, sub-adult and adult), sex (categorical; male vs. female), season (categorical: spring, summer, fall and winter) and abortion with infection were analyzed by Chi-square test and logistic regression. Differences were considered statistically significant when p < 0.05.

Results

Two hundred and three out of 262 sera tested (77.48%, 95% CI: 72.48 - 82.48%) were positive to BTV antibodies. In a total number of 262 samples, there were 208(79.38%) ewes and 54(20.60%) rams, as the Table 1 shows 175 (84.10%) of the ewes and 28 (51.80%) of the rams had antibodies against BTV. Statistically significant differences were evident between sexes ($\chi^2 = 23.79$, df = 1, p < 0.001). The odds of observation of infection in ewe in comparison with ram was 4.92(95.00% CI: 2.57 - 9.44). There was no statistically significant difference between infection and recent abortion, so that 56 from 65 of recently

aborted ewes and 116 from 136 of normally delivered ewes were seropositive to BTV ($\chi^2 = 0.02$, df = 1, p > 0.05). The odds of infection in recently aborted ewes in comparison with normally delivered ewes was 1.07 (95.00% CI: 0.46 - 2.51).

Differences between age classes were also observed (χ^2 = 49.24, df = 2, p < 0.001). The odds of infection in adult animals in comparison with juveniles was 12.51(95.00% CI: 5.15 - 30.36), sub-adult animals in comparison with juveniles was 1.86(95.00% CI: 0.77 - 4.48) and adult animals in comparison with sub-adult was 6.72(95.00% CI: 3.27 - 13.79). No statistically significant differences were observed in BTV seroprevalence within seasons (χ^2 =0.34, df = 3, p > 0.05), (Table 1).

Discussion

Three classifications of BTV status (BTV free zones, BTV seasonally free zones, and BTV infected zones), that affect transportation and free trade of ruminants have been defined. This study has shown that Kohgiluyeh and Boyer-Ahmad province is considered a BTV infected zone, with BTV infection being highly widespread (77.50%) in this province.

A seroprevalence (34.70%) of BTV infection has been reported in sheep flocks in West Azarbaijan, Iran. In that survey, 172 of 184 flocks were BTV seropositive sheep (93.50%).15 The higher seroprevalence obtained in our study compared to the result of Shoorijeh et al. could be related to spatial and temperature variations. 14 Spatial variations observed in seroprevalence among areas may also be due to differences in the distribution of Culicoides vectors.^{8,16} In the case of temperature, West Azarbaijan province is generally colder than Kohgiluyeh and Boyer-Ahmad areas and this low temperature can affect the existence of colicoides vector that are not able to live at low temperatures. This explanation can be a reason why lower seroprevalence was seen in West Azarbaijan compared to Kohgiluyeh and Boyer-Ahmad. 17,18 There is another hypothesis about high seroprevalence of BTV in this area. Pakistan is an eastern neighbor of Iran and high seroprevalance of the BTV infection (48.40%) is reported

in Pakistan.¹⁹ The large volume of animal trade between Iran and Pakistan, especially sheep, could be the reason of high seroprevalance of BTV infection in Iran.

Wild ruminants may play a role in the epidemiology of BTV and they could act as reservoirs in transmission and maintenance of the virus.^{20,21} The existence of wild ruminants in Kohgiluyeh and Boyer-Ahmad province could have an important influence on the evolution of infection in livestock in this province.

Sheep over one year old (sub-adults and adults) have significantly higher seroprevalences (p < 0.001) than juveniles in this study. This is not unexpected, because animals older than 1 year old are likely to have been exposed to the risk of infection for longer than juvenile animals.²² Also, higher seroprevalence among adult sheep was likely due to acquired immunity gained over multiple years of exposure to BTV throughout multiple BTV vector seasons.¹¹ The seroprevalence in females was higher than males that may be due to the effect of age and sample size.

Nomads in Kohgiluyeh and Boyer-Ahmad province migrate from cold to moderate areas in winter and from warm to moderate areas in summer to maintain stable weather (moderate weather) for their animals throughout the year. This migration and the associated stable weather conditions may be the reason for our results that showed ineffectiveness of season in seroprevalence status. Some of flocks in this study were migrating and those flocks also can be a source of infection of BTV in this province.²³

Temperature, international trade, geographic status of areas, wildlife characteristic and lifestyle of people are of important factors that all of them can influence prevalence of BTV in the area. Thus, BTV prevalence should be investigated from several perspectives in an area, however, these views are sometimes interrelated and should be taken into consideration altogether and not in isolation.

In conclusion, this study confirmed that the BTV infection existed in Kohgiluyeh and Boyer-Ahmad province. Since a vaccination program for BT is not established in Iran, a seropositive result indicates BT infection in the domestic populations. According to local weather conditions and facility of vector-borne transmission, prevention and control measures should be considered by health authorities.

Table 1. Prevalence against BTV antibodies in sheep from southwest in Kohgiluyeh and Boyer-Ahmad province, Iran.

Category	Groups	Positive	Negative	Total
Sex	Female	175(84.13%)	33(15.87%)	208(79.39%)
	Male	28(51.85%)	26(48.15%)	54(20.61%)
Abortion	Recently aborted	56(86.15%)	9(13.85%)	65(32.34%)
	Delivered normally	116(85.29%)	20(14.71%)	136(67.66%)
Age	Juvenile	13(43.33%)	17(56.67%)	30(11.45%)
	Sub-adult	37(58.73%)	26(41.27%)	63(24.05%)
	Adult	153(90.53%)	16(9.47%)	169(64.50%)
Season	Spring	45(75.00%)	15(25.00%)	60(22.90%)
	Summer	50(79.37%)	13(20.63%)	63(24.05%)
	Fall	49(77.78%)	14(22.22%)	63(24.05%)
	Winter	59(77.63%)	17(22.37%)	76(29.00%)

Acknowledgements

We gratefully acknowledge Shahid Chamran University of Ahvaz for supporting this study.

References

- 1. Mertens PPC, Maan S, Samuel A, et al. Orbivirus, Reoviridae. In: Fauquet CM, Mayo MA, Maniloff J, et al (Eds). Virus taxonomy, 8th report of the ICTV. London, UK: Elsevier Academic Press 2005; 466-483.
- 2. Mellor PS, Carpenter S, Harrup L, et al. Bluetongue in Europe and the Mediterranean basin: History of occurrence prior to 2006. Prev Vet Med 2008; 87:4-20.
- 3. Monath TP, Guirakhoo F. Orbivurses and Coltiviruses. In: Fields BN, Knipe DM, Howley PM (Eds). Fields Virology. Philadelphia, USA: Lippincott-Raven Publisher 1996; 1735-1766.
- 4. Koumbati M, Mangana O, Nomikou K, et al. Duration of bluetongue viraemia and serological responses in experimentally infected European breeds of sheep and goats. Vet Microbiol 1999; 64: 277-285.
- 5. Alexander GI, Alexander MP, St George TD. Bluetongueits impact on international trade in meat and livestock. In proceedings: The first south-east Asia and Pacific regional bluetongue symposium. Canberra, Australia. 1995. 254-258.
- 6. Mac Lachlan NJ. The pathogenesis and immunology of bluetongue virus infection of ruminants. Comp Immunol Microbiol Infect Dis 1994; 17: 197-206.
- 7. Purse BV, Mellor PS, Rogers D, et al. Climate change and the recent emergence of bluetongue in Europe. Nat Rev Microbiol 2005; 3: 171-181.
- 8. Mellor PS, Wittmann EJ. Bluetongue virus in the Mediterranean basin, 1998-2001. Vet J 2002; 16:20-37.
- 9. Gibbs EP, Greiner EC. The epidemiology of bluetongue. Comp Immunol Microbiol Infect Dis 1994; 17:1737-1753.
- 10. Tabachnick WJ. Culicoides and the global epidemiology of bluetongue virus infection. Vet Ital 2004; 40:145-150.
- 11. Mayo C. Prevalence and risk factors associated with bluetongue virus among Colorado sheep flocks. Master

- thesis. Colorado State University, Fort Collins, Colorado 2010; 14-15.
- 12. Smith BP. Large animal internal medicine. 4th ed. California, USA: Elsevier 2009; 777-787.
- 13. Afshar A, Thomas FC, Wright PF, et al. Comparison of competitive and indirect enzyme-linked immunosorbent assays for detection of bluetongue virus antibodies in serum and whole blood. J Clin Microbiol 1987; 25(9): 1705-1710.
- 14. World organization for animal health, OIE. Terrestrial manual article: Bluetongue 2008; chapter 8.3: 448.
- 15. Shoorijeh SJ, Ramin AG, MacLachlan NJ, et al. High seroprevalence of bluetongue virus infection in sheep flocks in West Azerbaijan, Iran. Comp Immunol Microbiol Infect Dis 2010; 33: 243-247.
- 16. Calvete C, Miranda MA, Estrada R, et al. Spatial distribution of Culicoides imicola, the main vector of bluetongue virus, in Spain. Vet Rec 2006; 158: 130-131.
- 17. MacLachlan NJ. Bluetongue: A review and global overview of the only OIE list a disease that is endemic in North America. In proceedings: 55th annual meeting of the American college of veterinary pathologists (ACVP) and 39th annual meeting of the American society of clinical pathology. Middleton, USA. 2004; 143.
- 18. Wilson A, Darpel K, Mellor PS. Where does bluetongue virus sleep in the winter? PLoS Biology 2008; 6:1612-1617.
- 19. Akhtar S, Djallem N, Shad G, et al. Bluetongue virus seropositivity in sheep flocks in North West Frontier province, Pakistan. Prev Vet Med 1997; 29: 293-298.
- 20. Gortázar C, Acevedo P, Ruiz-Fons F, et al. Disease risk and overabundance of game species. Eur J Wildl Res 2006; 52: 81-87.
- 21. García I, Napp S, Casal J, et al. Bluetongue epidemiology in wild ruminants from southern Spain. Eur J Wildl Res 2009; 55: 173-178.
- 22. Ward MP, Carpenter TE, Osburn BI. Host factors affecting seroprevalence of bluetongue virus infections of cattle. Am J Vet Res 1994; 55(7): 916-920.
- 23. Bhalodiya MB, Jhala MK. Seroepidemiological study of blue tongue virus using AB-ELISA. Indian Vet J 2002; 79: 1237-1240.