

Serological study of bovine herpes virus type 1 in dairy herds of Hamedan province, Iran

Aliasghar Bahari^{1*}, Jamal Gharekhani², Masoumeh Zandieh², Ali Sadeghi-Nasab¹, Hesameddin Akbarein³, Ahmad Karimi-Makhsous², Morteza Yavari¹

¹ Department of Clinical Sciences, Faculty of Paraveterinary Sciences, Bu-Ali Sina University, Hamedan, Iran; ² Iranian Veterinary Organization, Hamedan, Iran; ³ Department of Food Hygiene and Quality Control, Faculty of Veterinary Medicine, University of Tehran, Tehran, Iran.

Article Info	Abstract
<p>Article history:</p> <p>Received: 03 September 2012 Accepted: 16 February 2013 Available online: 15 June 2013</p> <p>Key words:</p> <p>BHV-1 Cattle ELISA Hamedan Iran</p>	<p>A cross-sectional study with a random cluster sampling design was carried out to estimate the seroprevalence of bovine herpesvirus type 1 (BHV-1) in non-vaccinated dairy herds in Hamedan province, west of Iran. Simple random sampling was used for selection of cattle in each herd. Informative data about each herd and selected animals were recorded by the farm manager in a provided questionnaire. Blood samples were collected from 492 animals in 41 industrial herds. A commercial indirect ELISA test was used to determine the seropositivity against BHV-1. The individual and herd seroprevalence for BHV-1 were 58.74% and 82.93%, respectively. The intra-herd prevalences were ranged from 16.70% to 100%. Geographical characteristics of Hamedan province may explain the high seroprevalence rates found in this study compared to those of others obtained from different parts of the country. The proportion of seropositive cows were increased with age ($p < 0.05$). Animals from large and moderate sized herds had higher odds of seropositivity than those of small size herds. These findings could be related to the presence of a considerable number of BHV-1 carriers in this region. The high herd and animal prevalence found in the present study suggested necessity of implementing an intensive control program for reducing BHV-1 infection rates.</p> <p style="text-align: right;">© 2013 Urmia University. All rights reserved.</p>

مطالعه سرولوژیک آلودگی با هرپس ویروس گاوی تیپ یک (BHV-1) در گاوداری های شیری استان همدان-ایران

چکیده

یک مطالعه مقطعی با طراحی نمونه گیری خوشه ای تصادفی برای تخمین شیوع سرمی آلودگی با هرپس ویروس گاوی تیپ یک در گاوهای شیری واکسینه نشده در گاوداری های صنعتی استان همدان در غرب ایران انجام شد. در هر گله، گاوها به صورت تصادفی ساده انتخاب شدند. اطلاعات مربوط به گله ها و گاوهای نمونه گیری شده از مدیر گاوداری اخذ و در پرسشنامه ای ثبت شد. از ۴۹۲ رأس گاو مربوط به ۴۱ گله نمونه خون تهیه شد. برای آزمایش نمونه های سرم از کیت تجارتي الایزای غیرمستقیم استفاده گردید. شیوع سرمی انفرادی و گله ای هرپس ویروس گاوی تیپ یک به ترتیب ۵۸/۷۴ درصد و ۸۲/۹۳ درصد بود. شیوع داخل گله ای نیز از ۱۶/۷ تا ۱۰۰ درصد متغیر بود. ویژگی جغرافیایی استان همدان می تواند توجیه کننده میزان بالای شیوع سرمی بدست آمده در این مطالعه در مقایسه با یافته های سایر مناطق کشور باشد. نسبت گاوهای سرم مثبت، با بالا رفتن سن افزایش یافت ($p < 0/05$). شانس سرم مثبت شدن گاوها در گله های بزرگ و متوسط بیش از گله های کوچک بود. این یافته ها می تواند به حضور تعداد قابل توجه حامل های هرپس ویروس گاوی تیپ یک در این منطقه ارتباط داشته باشد. شیوع بالای گله ای و انفرادی در این مطالعه، ضرورت نهادینه کردن یک برنامه کنترلی فشرده را برای کاهش میزان آلودگی با هرپس ویروس گاوی تیپ یک پیشنهاد می کند.

واژه های کلیدی: الایزای، ایران، گاو شیری، هرپس ویروس گاوی تیپ یک، همدان

*Correspondence:

Aliasghar Bahari. DVM, DVSc
Department of Clinical Sciences, Faculty of Paraveterinary Sciences, Bu-Ali Sina University, Hamedan, Iran.
E-mail: aliasghar.bahari@basu.ac.ir

Introduction

Infectious bovine rhinotracheitis/infectious pustular vulvovaginitis (IBR/IPV), caused by bovine herpes virus type 1 (BHV-1), is an important disease of domestic and wild cattle.¹ Clinical signs of BHV-1 infection include symptoms of inflammatory reactions in both respiratory and genital tracts, and abortion. A systemic disease affecting visceral organs may develop in young calves.² BHV-1 is a member of genus *Varicellovirus* in the subfamily *Alphaherpesvirinae*, which belongs to the *Herpesviridae* family. However, there is only one antigenic serotype of BHV-1 by conventional serological assays.³ Two subtypes of BHV-1.1 (respiratory subtype) and BHV-1.2 (respiratory and genital subtype) have been described for this virus, based on restriction enzyme cleavage patterns of viral DNA.¹

The BHV-1 infection is mainly transmitted via respiratory, ocular or genital secretions through direct contact between animals. However, the infection may also spread via fresh or frozen semen from infected bulls as well as contaminated equipments.¹ Following a primary infection with a field isolate or vaccination with an attenuated strain, BHV-1 can become latent. The latent BHV-1 infection can become reactivated and the animal may shed the virus following stimuli, for example transport, parturition and glucocorticoid therapy.⁴

BHV-1 infections can be diagnosed by serological tests that detect the virus, viral antigen or specific antibody and by nucleic acid-based tests that detect genomic DNA, nucleic acid hybridization and sequencing. The immune response to primary BHV-1 infection in experimentally infected cattle is characterized by the development of specific IgM and IgG antibodies at 7 days post inoculation. Bovine herpes virus type 1 infected cows are mainly detected by the presence of specific antibodies to the virus after acute phase and during latency. Both virus neutralization (VN) test and ELISA have been employed for detecting antibodies against BHV-1. The ELISA is a specific, sensitive and practical test for detection of antibodies and has advantages over the VN test.^{5,6} Moreover, several types of BHV-1 ELISA tests are commercially available and some of them can be used in conjugation with marker vaccines to detect infected cattle in vaccinated populations.⁶⁻⁸

The detection of latent BHV-1 infection in cattle is important for control programs and trade activities in dairy farms. Several studies have already demonstrated large different seroprevalence from 22.68% in the south⁹ to 48.90% in the northwest of Iran.¹⁰ There is no published data on BHV-1 prevalence in dairy herds located in western provinces of Iran. Therefore, the main purpose of the present study was to determine the seroprevalence of BHV-1 infection in dairy cattle herds of Hamedan province, Iran.

Materials and Methods

Study design. A cross-sectional study, with a random cluster sampling design, carried out in 41 industrial dairy herds of Hamedan province (situated in the middle of western Iran) between June and August 2009. The study area lies between longitudes 47° 34' to 49° 36' E and latitudes 33° 59' to 35° 48' N. Hamadan province is located in a temperate mountainous region with mean annual precipitation of 330 mm and mean annual temperature is 11.4 °C. The populations of cows in the tested farms were 50 to 660. Before sampling, the herds were divided to small (50-100 cows), medium (100-400 cows) and large (> 400 cows) units. No vaccination against BHV-1 was carried out in the farms; however, artificial insemination was applied in most of those herds. In each selected farm, individual cattle were chosen by random sampling method. The tested animals were divided to group A (between 6 months and 2 years), B (between 2 and 3 years), C (between 3 and 4 years) and D (more than 4 years), in order to study the association of age with seroprevalence of the virus. To avoid false positive results due to potentially presence of maternal antibodies, calves younger than six months age were excluded from the study.

Serological test. Four hundred ninety two blood samples were collected from coccygeal vein using plain vacutainer tubes, and transferred to the laboratory on ice. The samples were centrifuged at 2000 *g* for 10 min to obtain serum. Sera were stored in the labeled microtubes at -20 °C until analyzed. BHV-1 specific antibodies in the serum samples were detected using bovine rhinotracheitis virus (BHV-1) gE antibody ELISA test kit (HerdCheck® IDEXX Laboratories, Inc., The Netherlands) according to manufacturer's instruction. Optic density (OD) was read at 650 nm in a multi-well plate reader (ELx800, BioTek Inc., Winooski, VT, USA). According to the manufacturer, the sensitivity and specificity of the kit was 98.00%. The Serum samples with OD \geq 0.50 were considered as positive cases.

Data analysis. Considering the reported specificity of the used kit (98.00%), herd with zero or one seropositive animals was defined as negative and herds with at least 2 seropositive animal as positive. Herd size, within-herd prevalence, overall individual and herd prevalence were calculated on the basis of the number of tested animals. Data analysis of Chi-Square and Fisher's exact test were carried out by SPSS version 16.0 (SPSS Inc., Chicago, IL, USA). A *p* value of less than 0.05 was considered statistically significant.

Results

Seroprevalence of BHV-1 antibody in sampled animals was found to be 58.74% (289 of 492 sera) and according to presence of at least 2 seropositive animals in a herd, 34 herds (82.93%) were seropositive (Table 1).

Table 1. Seroprevalence of BHV-1 within examined herds and tested dairy cattle in Hamedan province, Iran (%).

Herd size*	No. of herds	No. of sero-positive herds	No. of samples	No. of sero-positives [†]	No. of sero-negatives [†]
Small	18	14	216	104 (48.15) ^a	112 (51.85) ^a
Medium	18	15	216	139 (64.35) ^b	77 (35.65) ^b
Large	5	5	60	46 (76.67) ^b	14 (23.33) ^b
Total	41	34	492	289 (58.74)	203 (41.26)

* Small: 50-100, medium: 100-400, large: > 400 dairy cows in herd.

[†]In each column, different letters show significant difference ($p < 0.05$).

The prevalence was ranged from 16.70 to 100% among seropositive herds in this study. All of 7 (17.10%) negative herds were from small and medium size herds (Table 1). Seroprevalence of BHV-1 within age groups was increased with age (Table 2). Based on age, the prevalence of seropositivity among group A was significantly different from other groups ($p < 0.05$). However, the prevalence rates between age groups of B, C and D showed no significant differences (Table 2).

Table 2. Distribution of BHV-1 seroprevalence in dairy cattle with different ages of Hamedan province, Iran (%).

Groups	Age	No. of samples	No. of sero-positives *	No. of sero-negatives *
A	6 months to 2 years	55	21 (38.18) ^a	34 (61.82) ^a
B	2 to 3 years	113	60 (53.10) ^b	53 (46.90) ^b
C	3 to 4 years	147	93 (63.27) ^b	54 (36.73) ^b
D	more than 4 years	177	115 (64.97) ^b	62 (35.03) ^b
Total		492	289 (58.74)	203 (41.26)

* In each column, different letters show significant difference ($p < 0.05$).

Discussion

The seroprevalence of BHV-1 in tested herds (82.93%) and tested animals (58.74%) found in this cross-sectional study was more than previously reported studies. Serological studies on the dairy cattle of different parts of Iran demonstrated 22.68% infection in Shiraz,⁹ 30.39% in Kerman,¹¹ 31.48% in Ahvaz,¹² 46.68% in Chaharmahal and Bakhtiari province¹³ and 48.9% in Urmia.¹⁰ It is reported that farms located in colder and higher altitude area are more prone to have experienced BHV infections.¹⁴ The climate and/or dairy cattle practice density in regions with higher prevalence is almost similar to Hamedan province from which obtained results were also similar. On the other hand, industrial dairy herd farming is not a common practice in low altitude areas of Iran which are often tropical and dry. The geographical variations may explain the differences between our results and low prevalence areas such as Shiraz, Kerman and Ahvaz. Infection prevalences in small herds are significantly less than moderate and large ones (Table 1). In our opinion, beside to lower natural exposure in small herds, biosecurity programs might be better implemented than moderate and large herds. The herd and animal BHV-1 prevalence in the current study did not differ greatly with those reported from countries that have no control program for BHV-1 infection.

The prevalence rates were 84.00% for dairy herds and 35.00% for dairy cows in Belgium,¹⁵ 50.0% in Germany,¹⁶ 80% for Hungary¹⁷ before start of their eradication programs and 61.00% in unvaccinated dairy herds of Italy.¹⁸ In 107 non-vaccinated herds in south west of England, mean seroprevalence of cattle and herds have been detected 42.50% and 43.00%, respectively.¹⁴ Considering herd size and management systems of dairy herds in each region or country, all the mentioned studies reported a moderate to high prevalence of infection prior to beginning an eradication program which may reflect the latency ability of the virus.

All breeds of cattle at any age are susceptible but the disease occurs most commonly in animals over 6 months old, probably because of their greater exposure (e.g. nasal exudate and coughed-up droplets, genital secretions, semen, fetal fluids and tissues and etc.) to the infective agent and loss maternal immunity.¹⁹ There was a positive relationship between age and proportion of seropositive cows in the present study. Similar findings were also reported by Boelaert *et al.*¹⁵ and Solis-Calderon *et al.*²⁰ Since vaccination against BHV-1 was not practiced in the tested dairy herds, seropositivity was probably due to their naturally greater exposure to the virus. This fact is clearly demonstrated by significant difference between group A and other age groups (Table 2). Antibodies against BHV-1 persist throughout the animal's life after a natural infection.¹ Moreover, the older animals are more likely to be exposed to natural sources of infection.²¹ Badiei *et al.* did not find any difference in the number of seropositive cows between age groups.⁹ They explained that their unexpectedly findings might be due to difference in herd size, cow rearing systems and animal keeping conditions.

Serologically positive cows which are latent carriers have the key roles in reserving and transmission of BHV-1 to susceptible animals.²² In agreement with Boelaert *et al.* and Solis-Calderon *et al.* the prevalence rate higher than 38.00% (Table 1) may indicate the naturally circulation of BHV-1 among animals.^{15,20} Moreover, the largest herds and older animals had highest seropositivity (Tables 1 and 2). This may show a high number of BHV-1 carriers in this herds. The method of isolation and slaughter of seropositive animals based on Danish and Swedish IBR control programs is inefficient for eradication of infection in countries with large herds or high seroprevalence of BHV-1. Several European countries have initiated control programs for eradication purposes based on using inactivated or live attenuated marker vaccines but no vaccine is able to prevent the infection and the establishment of latency.^{23,24} The marker vaccines used together with a serological detection of glycoprotein E-specific antibodies, allow differentiation of naturally infected cows from vaccinated animals.²⁵

In conclusion, the high prevalence in herds and animals found in this study suggested necessity of an intensive control program for reducing BHV-1 infection rates. Based on present findings, we recommend using marker vaccine

and serologically differentiation of naturally infected cows from vaccinated animals for eradication of IBR/IPV. Planned biosecurity measures are needed to control the epidemiological risk of infection due to the presence of BHV-1 latent carriers.

Acknowledgements

The authors thank to Dr. Pouya Zamani, Department of Animal Sciences of Bu-Ali Sina University, for performing a part of statistical analysis.

References

- Muyllkens B, Thiry J, Kirten P, et al. Bovine herpesvirus 1 infection and infectious bovine rhinotracheitis. *Vet Res* 2007; 38: 181-209.
- Diaz F. Manual of diagnostic tests and vaccines for terrestrial animals, 7th ed. Paris: Office International des Epizooties 2012: 1343.
- Hage JJ, Schukken YH, Schols H, et al. Transmission of bovine herpesvirus 1 within and between herds on an island with a BHV1 control programme. *Epidemiol Infect* 2003; 130: 541-552.
- Pastoret PP, Thiry E, Brochier B, et al. Bovine herpesvirus 1 infection of cattle: pathogenesis, latency, consequences of latency. *Ann Vet Res* 1982; 13: 221-235.
- Kaashoek MJ, Moerman A, Madić J, et al. A conventionally attenuated glycoprotein E-negative strain of bovine herpesvirus type 1 is an efficacious and safe vaccine. *Vaccine* 1994; 12: 439-444.
- Van Oirschot JT, Kaashoek MJ, Maris-Veldhuis MA, et al. An enzyme-linked immunosorbent assay to detect antibodies against glycoprotein gE of bovine herpesvirus 1 allows differentiation between infected and vaccinated cattle. *J Virol Methods* 1997; 67: 23-34.
- Lehmann D, Sodoyer R, Leterme S, et al. Improvement of serological discrimination between herpesvirus-infected animals and animals vaccinated with marker vaccines. *Vet Microbiol* 2002; 86: 59-68.
- Mars MH, De Jong MCM, Franken P, et al. Efficacy of a live glycoprotein E-negative bovine herpesvirus 1 vaccine in cattle in the field. *Vaccine* 2001; 19: 1924-1930.
- Badiee K, Ghane M, Mostaghni K. Seroprevalence of bovine herpes virus type 1 in the industrial dairy cattle herds in suburb of Shiraz-Iran. *Aust J Basic and Appl Sci* 2010; 4(10): 4650-4654.
- Mahmodian AR, Dalir-Naghade B, Rahmati R. Serological study of BHV-1 in Urmia cattle by ELISA. In Proceedings: The third Convention of Iranian Veterinary Clinician. Mashhad, Iran. 2002; 355.
- Sakhaee E, Khalili M, Kazemini S. Serological study of bovine viral respiratory diseases in dairy herds in Kerman province, Iran. *Iranian J Vet Res* 2009; 10:49-53.
- Haji Hajikolaei MR, Seyfiabad Shapouri MR. Sero-epidemiological study of bovine herpesvirus 1 (BHV-1) infection in cattle in Ahvaz. *Iranian Vet J* 2006; 2: 23-31.
- Hemmat Zade F, Momtaz H, Tajbakhsh E, et al. A serological survey on bovine herpesvirus 1 (BHV-1) in ChaharMahal and Bakhtiari province, Iran. *Pajouhesh and Sazandegi* 2002; 15: 38-43.
- Woodbine KA, Medley GF, Moore SJ, et al. A four year longitudinal seroepidemiological study of bovine herpesvirus type-1 (BHV-1) in adult cattle in 107 unvaccinated herds in south west England. *BMC Vet Res* 2009; 5, 5. doi: 10.1186/1746-6148-5-5.
- Boelaert F, Biront P, Soumare B et al. Prevalence of bovine herpesvirus-1 in the Belgian cattle population. *Prev Vet Med* 2000; 45:285-295.
- Teuffert J. The national IBR/IPV- eradication program in Germany. Achievements and problems. In Proceedings: BHV-1 eradication symposium. Berlin, Germany 2006; 14.
- Pálfi V, Földi J. Experiences on BHV-1 eradication in Hungary. In Proceedings: BHV-1 eradication Symposium. Berlin, Germany 2006; 15.
- Cavinari S. Epidemiological data on IBR in Italy and experience of control in the field. In Proceedings: BHV-1 eradication Symposium. Berlin, Germany 2006; 8-9.
- Radostits OM, Gay CC, Hinchcliff KW, et al. *Veterinary Medicine*, 10th ed. London, UK: WB Saunders 2007: 1349-1361.
- Solis-Calderon JJ, Segura-Correa VM, Segura-Correa JC, et al. Seroprevalence of and risk factors for infectious bovine rhinotracheitis in beef cattle herds of Yucatan, Mexico. *Prev Vet Med* 2003; 57:199-208.
- Nardelli S, Marangon S, Dalla poza M, et al. Bovine herpesvirus 1 (BHV-1) seroprevalence in the breeding cattle population of the Veneto region: prospects for the implementation of a control programme. *J Vet Med B* 1999; 46: 735-740.
- Hage JJ, Schukken YH, Barkema HW, et al. Population dynamics of bovine herpesvirus 1 infection in a dairy herd. *Vet Microbiol* 1996; 53:169-180.
- Wizigmann G. Qualifying and monitoring of IBR-free herds in Bavaria (Germany). Maastricht, June 1997. Available at: www.animalhealthservice.nl/pages/staff/litlijst/ibr/wizig.htm. Accessed Nov 13, 2002.
- DeWit JJ, Hage JJ, Brinkhof J, et al. A comparative study of serological tests for use in the bovine Herpesvirus 1 eradication programme in The Netherlands. *Vet Microbiol* 1998; 61:153-163.
- van Drunen Littel-van den Hurk S, Van Donkersgoed J, Kowalski J et al. A subunit gIV vaccine, produced by transfected mammalian cells in culture, induces mucosal immunity against bovine herpesvirus-1 in cattle. *Vaccine* 1994; 12 1295-1302.